

Fossils provide better estimates of ancestral body size than do extant taxa in fishes

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Abstract

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The use of fossils in studies of character evolution is an active area of research. Characters from fossils have been viewed as less informative or more subjective than comparable information from extant taxa. However, fossils are often the only known representatives of many higher taxa, including some of the earliest forms, and have been important in determining character polarity and filling morphological gaps. Here we evaluate the influence of fossils on the interpretation of character evolution by comparing estimates of ancestral body size in fishes (non-tetrapod craniates) from two large and previously unpublished datasets; a palaeontological dataset representing all principal clades from throughout the Phanerozoic, and a macroecological dataset for all 515 families of living (Recent) fishes. Ancestral size was estimated from phylogenetically based (i.e. parsimony) optimization methods. Ancestral size estimates obtained from analysis of extant fish families are five to eight times larger than estimates using fossil members of the same higher taxa. These disparities arise from differential survival of large-bodied members of early branching lineages, and are not statistical or taphonomic artefacts. Estimates of ancestral size obtained from a limited but judicious selection of fossil fish taxa are more accurate than estimates from a complete dataset of extant fishes.

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Introduction

The vast majority (> 99%) of species that have ever existed are now extinct (Simpson 1952) and whole branches of the tree of life are known only from fossil forms (e.g. trilobites, placoderms, plesiosaurs). Consequently, fossils represent a unique resource for evolutionary studies. However, interpreting the morphology of fossil taxa is regarded as more subjective and less informative than data derived from living members of the same higher taxon (Patterson 1981; Ax 1987). In particular, the morphology of extant taxa can be studied in greater detail than in fossils, including aspects of soft anatomy, and usually using larger sample sizes. By contrast, and despite occasionally exceptional preservation,

fossils are often fragmentary and exhibit large amounts of non-randomly distributed missing data (Wiens 2006). All of these sources of error contribute to uncertainty about the phenotypes and phylogenetic positions of fossil taxa (Gauthier *et al.* 1988; Wilkinson 2003).

Considering even limited sample sizes and problems associated with preservation of individuals, fossils can still provide irreplaceable information regarding the tempo and mode of character evolution. Fossils are often the only exemplars from the earliest radiations in many higher taxa, providing critical information in determining character polarity (Conway Morris 1993; Budd and Jensen 2000; Briggs and Fortey 2005). Because the taxa they represent are often extinct, fossils may represent taxa on shorter genealogical

branches than their living relatives, and as such are more likely to preserve less derived character states. Fossils often sample lineages closer in time to relatively deep splitting events, and frequently display character state combinations not observed among extant forms (Gauthier *et al.* 1988; Donoghue *et al.* 1989; Wilson 1992; Santini and Tyler 2004). Because of the added information from extinct lineages, inclusion of fossils in phylogenetic analyses has substantially improved understanding of phylogeny (O’Leary 1999; Gatesy and O’Leary 2001), character state evolution (O’Leary 2001) and phylogenetic trends (Finarelli and Flynn 2006; Cobbett *et al.* 2007).

Body size is among the most easily acquired and directly comparable attributes of organisms for which reliable estimates may be obtained from the fossil record (Stanley 1998). Adult body size is also a central feature of organismal design, imposing constraints on many aspects of life history, especially critical scaling functions related to growth, metabolism and fecundity (Peters 1983; Schmidt-Nielsen 1984; Haldane 1985; McNab 2002). Consequently, understanding the evolution of body size can provide insights into the diversification of biological form and function. Changes in body size can have allometric effects on morphology, physiology, behaviour (e.g. activity patterns, thermoregulation), and are widely used as an adaptation to novel physical (e.g. temperature extremes, hypoxia, desiccation) and biotic (e.g. predation, competition) environmental parameters (Hutchinson and MacArthur 1959; Strathdee and Bale 1998; Burness *et al.* 2001; Leaper *et al.* 2001). Importantly, many features associated with body size transcend the particularities of taxonomic design, and as such often exhibit repeated patterns of evolution (Wake 1991; Mabee 2000; Bird and Mabee 2002; Mabee 2002).

Fishes (non-tetrapod craniates) provide numerous examples of taxa and circumstances in which to test theories on the evolution of continuous traits such as size (Albert *et al.* 1999; Knouft and Page 2003). Marine and freshwater fishes represent the largest component of contemporary vertebrate diversity, including more than 50% of all living vertebrate species, and inhabiting most of the Earth’s aquatic habitats and geographical regions. Fishes also have a rich palaeontological record from throughout the Phanerozoic, with fossil taxa ranging in body size over more than three orders of magnitude. Moreover, recent discoveries of early Palaeozoic fossils have greatly expanded our knowledge of early vertebrate diversity and phylogeny (see Mallatt and Chen 2003; Shu *et al.* 2003; Janvier *et al.* 2006; references therein).

To better understand the importance of fossil data in documenting patterns of diversification in relation to body size, we compare estimates of ancestral body size in fishes using phylogenetically based methods of character state optimization applied to two new (previously unpublished) and large datasets. The first is a palaeontological dataset representing all principal clades of non-tetrapod craniates from throughout the Phanerozoic, including exemplars of all the early

Ordovician radiations (*c.* 488 Ma; Long 1995; Janvier 1996). The second dataset is a compilation of mean body size for all 515 families of living (Recent) fishes using data from FishBase (Froese and Pauly 2005). A main conclusion is that estimates of ancestral body size obtained from a limited but judicious selection of fossil taxa are more accurate than estimates from an (almost) complete dataset of all extant fishes. The results invite caution when interpreting the conclusions of character state optimization studies based on examination of extant taxa alone. These limitations persist even when the terminal taxa represent a complete (or almost complete) sampling of the living biota, and when they have been analysed in a robust phylogenetic context. These results are a reminder that patterns of organismic diversification arise from the processes of both speciation and extinction, and that only study of fossils allows the sampling of character states in extinct clades.

Materials and methods

Body size was measured as total length (TL) in centimetres from the tip of the snout to the posterior margin of the caudal fin directly from specimens, published photographs or reconstructions of articulated specimens in primary sources. Appendix 1 presents maximum recorded total length and stratigraphic data for 465 fish species, including 425 species known only as fossils and 40 species in clades for which fossils with reliable size data are lacking (e.g. *Myxine*; *Huso*). Appendix 2 presents statistical measures of size (average total length in cm), size variation (standard deviation, skew, kurtosis), and species richness (N) for all 515 recognized extant fish families, based on data for 24 259 species from FishBase. The taxonomic compositions of these two datasets are summarized in Tables 1 and 2, respectively. Total length is highly correlated with other measures of overall size, including maximum body weight, and size and age to first reproduction (Froese and Binohlan 2000). Among fishes, body mass in grams (g) may be estimated from total length in cm from the empirical equation: $g = 0.0217 TL^{2.861}$ (Fig. 1).

The fossil taxa included in this analysis were selected to maximize representation of phylogenetically basal craniate lineages (*sensu* Janvier 1996), and include a thorough sampling of higher fish taxa for which reliable estimates of size are currently available. Taxon sampling followed the basal exemplar approach which maximizes representation of phylogenetically basal clades (Prendini 2001; Prendini and Wheeler 2004). The basal-exemplar approach is less sensitive to preservational biases than stratigraphically based taxon-counting methods (Lane *et al.* 2005). This sampling strategy produced a fossil dataset that is broadly representative of the fossil record of fishes as a whole, including members of more than half (51%) of all the 324 fish families known only as fossils (Benton 1993), 26% (164 of 622) of all fish families, living and extinct, known as fossils, and 68% (71 of 105) of all non-teleost actinopterygian genera known only as fossils. Fossil

Table 1 Taxonomic summary of the fossil fish database

Clade	Fossil*	Extant†	Total	Total (%)
Cephalochordata	3	1	4	0.86
Yunnanozoa	2	0	2	0.43
Hyperotreti	3	7	10	2.15
Myllokunmingiida	2	0	2	0.43
Hyperoartia	5	5	10	2.15
Pteraspidomorphi	25	0	25	5.38
Thelodonti	8	0	8	1.72
Anaspida	6	0	6	1.29
Galeaspida	7	0	7	1.51
Pituriaspida	1	0	1	0.22
Osteostraci	15	0	15	3.23
Furcacaudiformes	2	0	2	0.43
Placodermi	39	0	39	8.39
Chondrichthyes	91	5	96	20.65
Acanthodii	17	0	17	3.66
Sarcopterygii	38	4	42	9.03
Actinopterygii	161	18	179	38.49
Total	425	40	465	100.00

*Taxa known only as fossils. †Extant taxa for clades lacking fossils with reliable size data (e.g. *Myxine*; *Huso*). Data are maximum recorded total length (cm), geological age (Epoch or Series), and phylogenetic position, from multiple sources (see text for explanation).

Table 2 Taxonomic summary of the extant fish species database. Data are maximum recorded total length (cm) and taxonomic affiliation for more than 24 000 species from Froese and Pauly (2005)

Clade	Orders	Families	Species	Total (%)
Hyperotreti	1	1	69	0.28
Hyperoartia	1	2	40	0.16
Chondrichthyes	13	46	826	3.40
Sarcopterygii	3	4	10	0.04
Actinopterygii	45	462	23 314	96.10
Total	63	515	24 259	100.00

species were dated to epoch or series (e.g. Upper Devonian, Palaeocene) with geological dates from Gradstein *et al.* (2004). Conodonts were excluded from analysis due to uncertainties in body size and detailed phylogenetic information (Donoghue and Sansom 2002; Janvier 2003; Dong *et al.* 2005; Northcutt 2005; Wickstrom & Donoghue 2005). Hyperoartia data are from Janvier and Lund (1983), Gess *et al.* (2006) and Janvier *et al.* (2006). Triassic neoselachians are known only from teeth and were excluded from analysis (Underwood 2006). Sarcopterygian data are largely from Clouthier and Forey (1991), Cloutier 1996, Cloutier and Ahlberg (1996) and Clouthier (1997). Actinopterygian data are largely from Coates (1998), Dietze (2000), Arratia (1997, 1999), Arratia and Cloutier (2002), Arratia and Clouthier (2004), Lund (2000) and Friedman and Blom (2006). Carboniferous actinopterygians are not considered

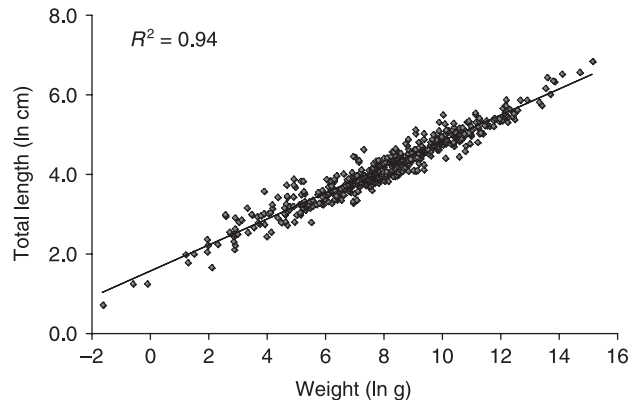


Fig. 1—Length and weight are significantly correlated in fishes. Data are maximum recorded total length (cm) and mass (g) for 517 extant fish species. The slope of the regression ($m = 0.3267$) on this log-log plot is close to the theoretically expected value 0.33 (i.e. $\text{mass} = \text{length}^3$). Size data from FishBase (Froese and Pauly 2005).

to be closely related to crown Actinopteri giving a long branch (*c.* 371–301 Ma).

The taxa examined provide sufficiently broad temporal (10^7 – 10^8 MY) and taxonomic (10^2 – 10^4 species) scopes to avoid non-random sampling errors arising from community assembly processes, convergent evolution or investigator bias (Ackerly 2000; Pollock *et al.* 2002). Due to the limited number of known fossils closely related to certain extant basal fish clades, size data for 13 terminal taxa are presented as an average of extant species from FishBase. These clades include the seven extant myxiniform genera, the two extant petromyzontiform genera, one extant dipnoan (*Protopterus* with six spp.), and five extant actinopterygians (*Polypterus*, *Acipenser*, *Scaphyrinchus*, *Psuedoscaphyrinchus*, *Clupea*). Taxa for which morphologically mature specimens are not known were excluded, as were taxa for which adult body size cannot be reliably estimated from known fossilized fragments (e.g. †*Polymerolepis margaritifera*, †*Lophosteus* sp.). Mature specimens are recognized by osteological criteria when available, that is, the shape of bones in the sphenoid and palatoquadrate regions of the skull, and the scapulocoracoid region of the pectoral girdle (Arratia 1997). Size of some Palaeozoic forms was estimated from large body fragments (e.g. †*Andinaspis suarezorum*, †*Pituriaspis doylei*; Janvier, pers. comm.).

Composite tree topologies were constructed from literature sources. The phylogeny of principal craniate clades (i.e. with initial radiations during the Lower Ordovician, *c.* 488–472 Ma; Fig. 2), of 465 fossil species, and of the 515 extant fish families (Fig. 3), largely follow Janvier (1996, 2003) and Long (1995), and references therein, with certain emendations noted by taxon in Appendix 1. These sources were used to construct a tree topology for fossil fishes with 843 branches and 86 polytomies, or a tree that is *c.* 91% resolved.

†extinct taxa known only as fossils.

Fig. 2—Interrelationships and stratigraphic ranges of the principal craniate clades (i.e. Ordovician radiations *c.* 488 Ma (Long 1995; Janvier 1996; Table 1). Thick lines represent known stratigraphic ranges. Extant craniates represent just five of the 14 principal craniate clades. In terms of numbers of clades and species, Actinopterygii (ray-finned fishes) dominates the marine ichthyofauna from the Carboniferous (*c.* 363 Ma) to the Recent, the global freshwater ichthyofauna from the Upper Cretaceous (*c.* 100 Ma) to the Recent, and includes 96.1% of living fish species.

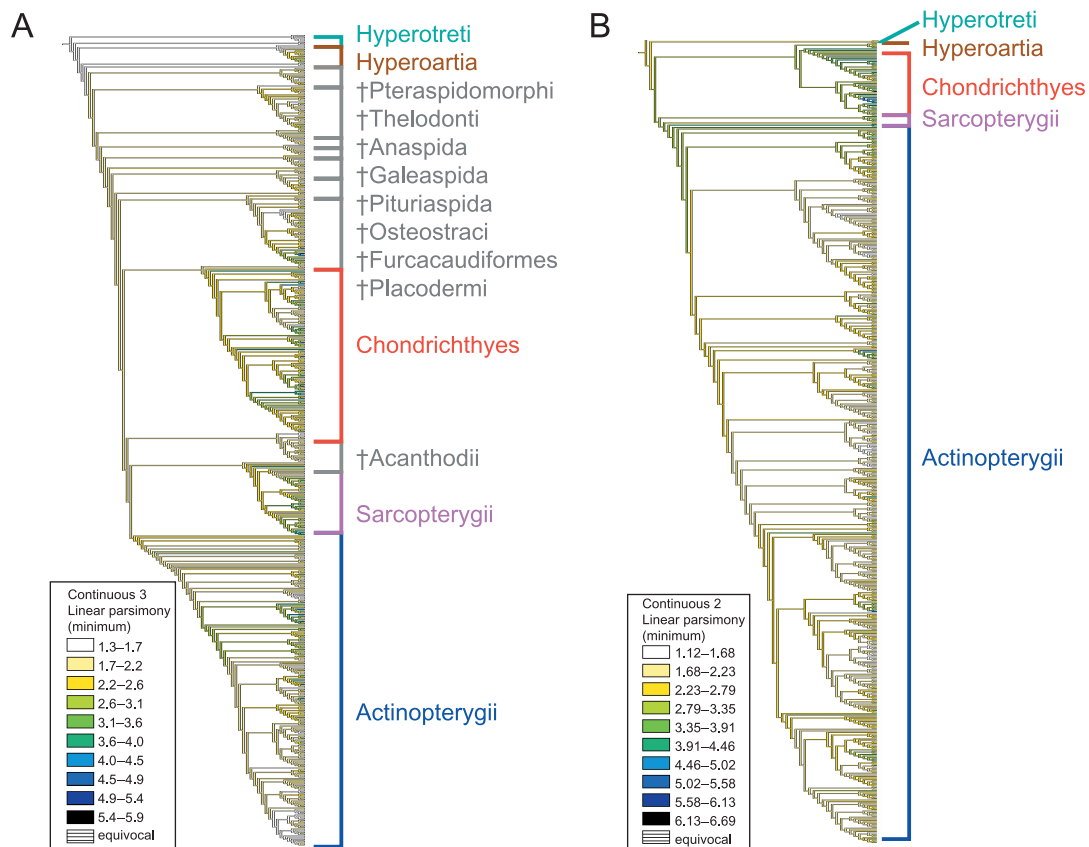
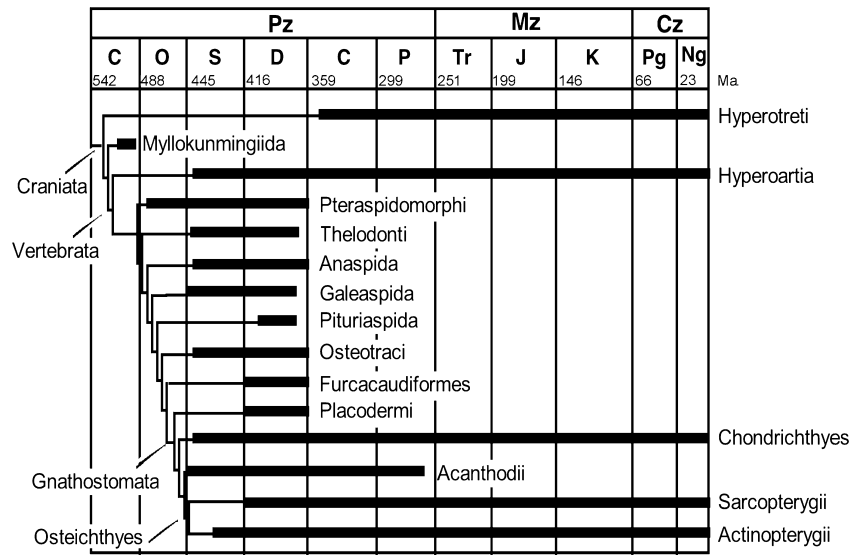


Fig. 3—Phylogenetic hypotheses of fish taxa with size-change optimized at all internal tree branches using Linear Parsimony (LP). —**A.** 465 fossil species, representing all 14 principal craniate clades (Ordovician radiations), with size optimized at 926 branches. —**B.** 515 extant fish families representing five principal craniate clades, with size optimized at 1031 branches. Tree topologies from Long (1995), Janvier (1996), Appendix 1 and references therein. Names of extinct clades (†) in grey font; extant clades in coloured fonts. Size data in cm log transformed before analyses. Branch lengths in MY estimated from stratigraphic data.

Two chordate outgroup taxa were used to root the size optimizations: Cephalochordata and Yunnanozoa (Mallatt and Chen 2003). The phylogenetic positions of †*Haikouichthys* as a non-craniate deuterostome follows Shu (2003).

Linear and least squared parsimony (LSP) methods were employed to optimize ancestral size using the MESQUITE v.1.06 software package (Maddison & Maddison 2006). Linear Parsimony (LP) minimizes the total amount of trait change along tree branches such that the cost of a change from state x to y is $|x - y|$ (Swofford and Maddison 1987). LSP, also referred to as Squared-Change Parsimony, follows a Brownian motion model of evolutionary change in which the cost of a change from state x to state y is $(x - y)^2$ (Maddison 1991). LP differs from LSP and model-based (i.e. Bayesian and Likelihood) approaches to character state optimization in that it permits the reconstruction of discontinuous events, or of large changes in trait values (Butler and Losos 1997; Pagel 1999). Although evolutionary change is often considered as gradual, large differences in trait values between internal tree nodes may result from a variety of real biological processes, including punctuated evolution (Pagel *et al.* 2006) or extinction of taxa with intermediate trait values (Butler and Losos 1997; Albert *et al.* 1998). LP also permits the reconstruction of ambiguous ancestral state values when data are insufficient to provide an unambiguous resolution. Nevertheless, estimates of mean size among fossil fishes per epoch using LP and LSP are significantly correlated ($P < 0.0001$; Fig. 4). All ancestral reconstruction methods assume that trait evolution is conservative enough for node reconstruction techniques to be useful, even in the face of large standard errors (Polly 2001).

Ancestral trait optimization was performed using 10 replicates on arbitrarily fully resolved trees using MACCLADE 4.07 (Maddison and Maddison 2005). The qualitative results of this analysis were similar in all replicates of arbitrary node resolution. Available methods of character state reconstruction are limited to estimating ancestral trait values from within the limits of those observed in terminal taxa. LP analysis may therefore perform poorly at detecting a consistent underlying trend like Cope's rule. The reader is referred to Albert (2006) for a discussion of the limits and assumptions of different ancestral trait reconstruction methods. Stratigraphic data of fossils were used to constrain minimum age estimates for internal tree branches (Benton and Donoghue 2007). Branch lengths were estimated from stratigraphic data from fossils following Benton (1993, 2005). Branch lengths were estimated as the absolute difference in MY between nodes. In several taxa known only from Recent organisms, branch lengths were estimated from biogeographical information among sister taxa (see Appendix 1).

Results and discussion

Ancestral size estimates obtained from analysis of the 515 extant fish families are five to eight times larger than esti-

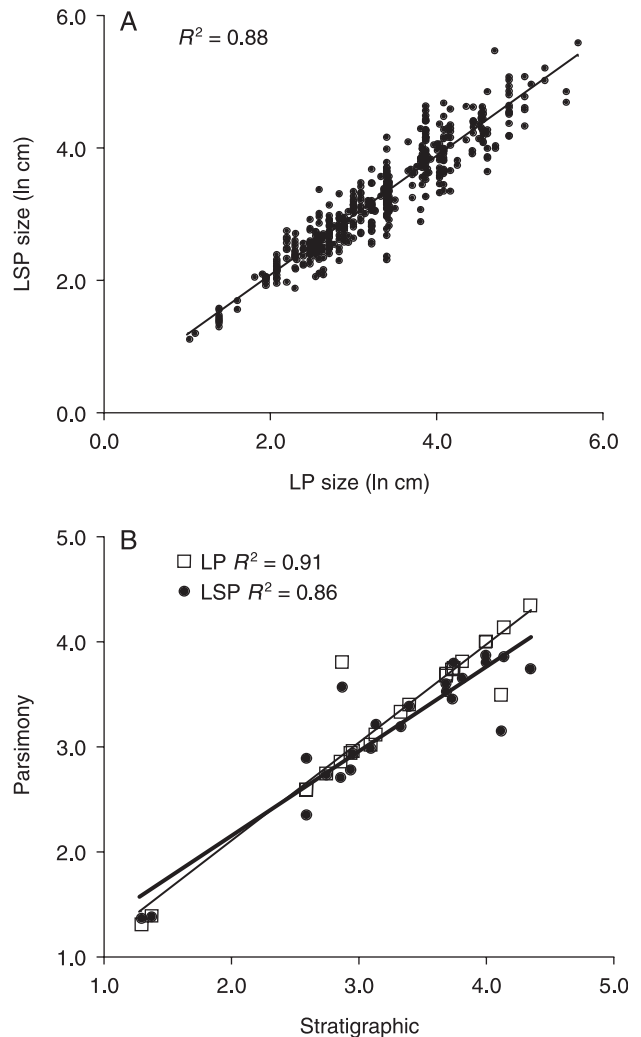


Fig. 4—Estimates of mean body size (ln cm) per epoch from phylogenetic optimization (LP and LSP) and stratigraphic (non-phylogenetic) methods. Stratigraphic estimates assessed directly as average log-transformed body size data of fossils per epoch. Phylogenetic estimates assessed as averages of interior node values per epoch using LP and LSP optimization on the phylogeny of fossil fishes (Fig. 3A). All R^2 values are significant at $P < 0.0001$. Note stratigraphic estimates are more highly correlated with LP than LSP, due to the averaging nature of squared-change optimization.

mates using fossil members of the same higher taxa (Fig. 5). This result is consistent for all of the 14 craniate clades with origins during the Lower Ordovician, including taxa with broad disparities in date of clade origin (c. 550–450 Ma), clade duration (c. 50–500 MY), body size at origin (c. 5–25 cm), habitat (i.e. marine, freshwater, euryhaline) and geography (i.e. tropical, extratropical, cosmopolitan). Plesiomorphic size estimates from the fossil dataset for Craniata, Hyperotreti, Vertebrata and Hyperartia are 5.0–8.0 cm, as compared with 45–50 cm from the extant dataset for these same taxa. Similarly, plesiomorphic size estimates from the

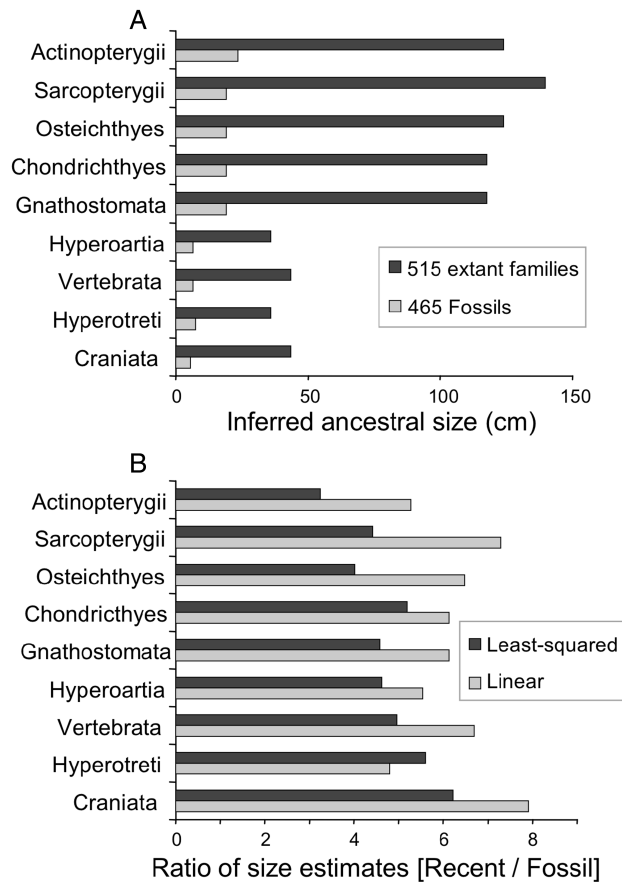


Fig. 5—Ancestral size estimates from analysis of extant fishes (515 extant families) are five to eight times larger than estimates using fossil members of the same higher taxa. —**A**. Inferred ancestral sizes using LP optimization. —**B**. Size estimates from extant (= Recent) vs. fossil members of the same higher taxa using LP and LSP optimization. Note the averaging effect of LSP results in somewhat less disparity in size estimate from Recent and fossil taxa.

fossil dataset for Gnathostomata, Chondrichthyes, Osteichthyes, Sarcopterygii and Actinopterygii are 20–25 cm, as compared with 120–140 cm from the extant dataset. Clearly there has been a strong filter on the size distribution of living taxa. A similar bias in the persistence of taxa with larger sizes has been observed in mammalian carnivores (Finarelli and Flynn 2006).

Which of the two datasets, palaeontological or contemporary provides more accurate estimates of size evolution among the principal craniate clades? Three features of craniate phylogeny and diversity suggest that information from fossils is more reliable for this purpose. First, the plesiomorphic size estimates of Craniata obtained from LP optimization of the fossil dataset are similar to the sizes (*c.* 3–5 cm) of closely related (fossil and extant) craniate outgroups (Mallatt and Chen 2003). Second, the extant diversity of fish clades represents only a fraction of the original (Ordovician) craniate

radiations (Webby *et al.* 2004), being limited to just five of the six clades that survived the Late Devonian crisis (*c.* 375 Ma; Fig. 2). This extinction event was a strong filter on the size as well as taxonomic composition of surviving fish faunas (Janvier 1996; McGhee 1996). Third, having persisted for longer periods of geological time, living members of a clade may be expected to have accrued on average more changes than lineages of the same clade cut short by extinction. As a result, fossil species often preserve plesiomorphic states with more fidelity than related extant species (Donoghue *et al.* 1989).

Why are size estimates derived from analysis of living fishes so much larger than those derived from fossil representatives of the same higher taxa? Such disparities could arise from systematic biases in methods used to assemble the fossil dataset or conduct the optimization analysis, reflecting taphonomic or statistical artefacts from size-selective preservation, recovery or identification of fossils. Alternatively, the disparities might arise from real differences in the evolutionary histories of taxa which have become extinct vs. those which have persisted to the Recent. Consideration of the available data suggests the different estimates of ancestral body size result from the persistence of phylogenetically basal taxa with large size among living fish clades. This result also reinforces the claim that early branching lineages do not necessarily retain primitive or ancestral traits (Crisp and Cook 2005).

The disparity in body size estimates from the fossil and extant datasets does not appear to be a taphonomic artefact arising from size-selective preservational biases. Size-related biases on the preservation, recovery and identification of fossils may provide potentially confounding signals in inferring size evolution from palaeontological data (Barton and Wilson 2005; Northwood 2005). Large specimens are more subject to disarticulation and dispersal through hydrodynamic transport and physical wear through abrasion (Long and Langer 1995; Butler and Schroeder 1998; Butler 2004). As a result, large fishes are less likely to be preserved as intact skeletons, and preserved isolated elements are less likely to be recovered and identified, thus hindering accurate estimates of body size. In this regard, Phanerozoic escalation of predation and bioturbation rates (Vermeij 1994) could influence long-term trends in the size-frequency distributions of fossil fishes through time. On the other hand, larger skeletal elements are usually more robust, more resistant to abrasion, and often have more readily observed diagnostic morphologies. As a result they are more likely to be preserved, recovered and correctly identified (Behrensmeier 1978; Kidwell and Flessa 1995; Alroy 2000). Indeed large-bodied taxa are better represented in many vertebrate palaeofaunas (Cooper *et al.* 2006; Valentine *et al.* 2006), and may actually serve to inflate perceptions of trends to larger size. The aggregate effect of these confounding taphonomic influences on size evolution remains poorly understood (Madin *et al.* 2006).

Global patterns of diversity may also reflect variation in the nature of the fossil record and fossil bearing sediments (Alroy

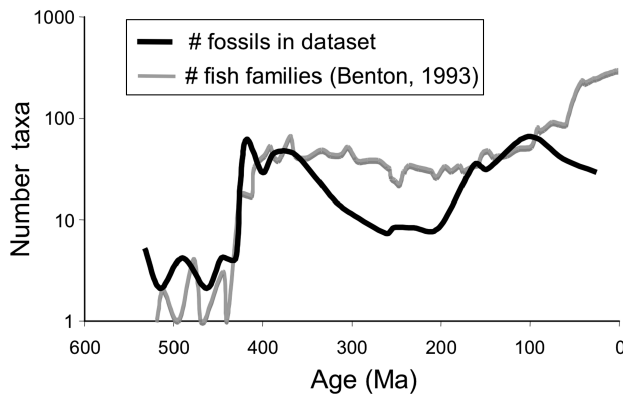


Fig. 6—Numbers of taxa per epoch in the fossil fish dataset (Appendix 1). The limited number of Late Palaeozoic (c. 318–251 Ma) fossils reflects a major trough in documented ichthyofaunas from the Pennsylvanian to Permian (Hurley *et al.* 2007). Family level diversity for 645 fish families from Benton (1993, 2005).

et al. 2001). For example, the dearth of fossil fish taxa from the Pennsylvanian to the Permian (318–251 Ma; Fig. 6) corresponds to a major trough in fish diversity and documented ichthyofaunas known from this interval (Sepkoski 2002; Hurley *et al.* 2007). Among stratigraphic intervals examined (Table 1), there are no significant correlations between mean body size and species richness or the proportion of articulated skeletons ($P > 0.1$). Madin *et al.* (2006) found that escalatory trends did not drive Phanerozoic macroevolutionary patterns in a large dataset of fossil benthic marine invertebrates. Similarly, body size has not been found to be correlated with other long-term geological trends, for example, sedimentary rock volume (Peters and Foote 2001; Crampton *et al.* 2003), bioturbation rates (Crimes and Droser 1992; Vermeij 1993) or mean size of top predators (Janvier 1996; Twitchett *et al.* 2005). In combination we take these results as evidence that taphonomic effects have not been the primary factor influencing the assessment of size distributions of fossil fishes over the Phanerozoic.

If, as predicted by Cope's rule, there was a persistent and general tendency to increase body size within lineages, ancestral size estimates obtained from analysis of terminal (fossil or extant) taxa would be systematically overestimated (Stanley 1973; Polly 1998; Hone and Benton 2005). For example, estimates from terminal taxa are limited to the range of values observed, and are not capable of estimating ancestral values smaller than that of the smallest terminal taxon. This overestimate in the value of internal tree nodes would arise regardless of optimization method used (i.e. LP vs. LSP). However, among 23 Phanerozoic epochs, estimates of internal node values from LP and LSP approaches are significantly correlated ($P < 0.001$) with those of a direct stratigraphic approach that does not use phylogenetic methods (Fig. 4). In other words, the principal qualitative results

of this study are similar regardless of the parsimony-based optimization approach employed.

The phylogenetic distribution of body size among living fishes strongly suggests a history in which the surviving members of basal taxa attain much larger sizes than did their fossil relatives. Among the principal craniate clades that emerged during the Ordovician and which have survived to the Recent, in all cases the living representatives are substantially larger than are the earliest fossils. To cite some examples, extant hagfishes (Myxiniiformes, avg. 51 cm, $n = 69$ species) are larger than the Pennsylvanian †*Myxinkela siroka* (7 cm) or †*Myxineides gononorum* (15 cm), extant lampreys (Petromyzontiformes, avg. 31 cm, $n = 40$ species) are larger than the Mississippian †*Hardistiella montanensis* (10 cm) or Pennsylvanian †*Mayomyzon pieckoensis* (6 cm), extant heterodontiform sharks (avg. 118 cm, $n = 8$ species) are larger than the Jurassic †*Heterodontus falcifer* (28 cm) or †*Paracestracion zitteli* (15 cm), extant coelacanth (avg. 154 cm, $n = 2$ species) are larger than the Middle Devonian †*Miguashaia bureaui* (50 cm) or Upper Devonian †*Diplurus newarki* (25 cm), extant lungfishes (Dipnomorpha, avg. 111 cm, $n = 8$ species) are larger than the Upper Devonian †*Rhinodipterus ulrichi* (28 cm), and extant non-teleost actinopterygians (avg. 165 cm, $n = 53$ species) are larger than Palaeozoic actinopterygians (e.g. Lower Devonian †*Dialipina salgueiroensis* at 25 cm, Middle Devonian †*Cheirolepis tralii* at 25 cm, Middle Devonian †*Stegotrachelys finlayi* at 10 cm, or Middle Devonian †*Moythomasia nitida* at about 10 cm). Some Palaeozoic actinopterygians did attain somewhat larger sizes (although not approaching modern values), especially during the Middle Devonian (e.g. †*Cheirolepis canadensis*, 55 cm) and Upper Devonian (e.g. †*Howqualepis rostridens*, 95 cm). Large size in these taxa is apparently derived (Lund and Poplin 2002; Arratia and Cloutier 2002; Arratia and Cloutier 2004; Friedman and Blom 2006).

To summarize, the available information pertaining to body size and phylogeny among the principal clades of fishes indicates differential survival of large-bodied members of early branching lineages. It is important to note these results pertain to phylogenetic patterns only, and do not directly address potential underlying evolutionary processes. In other words, we were not able to reject hypotheses of long-term anagenetic change (e.g. Cope's rule; Hone and Benton 2005), or of the effects of body size on relative rates of diversification (Brown 1999; Gillooly *et al.* 2001). The patterns of size evolution observed in fishes closely resemble those of other vertebrate clades examined to date with comparable taxonomic and temporal resolution (Gardezi and da Silva 1999; Laurin 2004; Smith *et al.* 2004; Webster *et al.* 2004).

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References

- Ackerly, D. D. 2000. Taxon sampling, correlated evolution, and independent contrasts. – *Evolution* 54: 1480–1492.
- Albert, J. S. 2006. Phylogenetic character reconstruction. In Kaas, J. H. (Ed.): *Evolution of Nervous Systems Volume 1: History of Ideas, Basic Concepts, and Developmental Mechanisms*, pp. 41–54. Academic Press, Oxford.
- Albert, J. S., Froese, R., Bauchot, R. and Ito, H. 1999. Diversity of brain size in fishes: Preliminary analysis of a database including 1174 species in 45 orders. In Seret, B. & Sire, J.Y. (Eds) *5th Indo-Pacific Fish Conference Proceedings*, pp. 647–656. Société Française d'Ichthyologie, Paris.
- Albert, J. S., Lannoo, M. J. and Yuri, T. 1998. Testing hypotheses of neural evolution in gymnotiform electric fishes using phylogenetic character data. – *Evolution* 52: 1760–1780.
- Alroy, J. 2000. New methods for quantifying macroevolutionary patterns and processes. – *Paleobiology* 26: 707–733.
- Alroy, J., Marshall, C. R., Bambach, R. K., Bezusko, K., Foote, M., Fursich, F. T. et al. 2001. Effects of sampling standardization on estimates of Phanerozoic marine diversification. – *Proceedings of the National Academy of Sciences of the United States of America* 98: 6261–6266.
- Arratia, G. 1997. Basal teleosts and teleostean phylogeny. – *Palaeo Ichthyologica* 7: 1–168.
- Arratia, G. 1999. The monophyly of the Teleostei and stem-group teleosts. In Arratia, G. and Schultz, H.-P. (Eds): *Mesozoic Fishes 2 – Systematics and Fossil Record*, pp. 265–334. Verlag Dr. Friedrich Pfeil, München.
- Arratia, G. and Clouthier, R. 2004. A new cheirolepidid fish from the Middle-Upper Devonian of Red Hill, Nevada, USA. In Arratia, G., Wilson, M. V. H. and Clouthier, R. (Eds): *Recent Advances in the Origin and Early Radiation of Vertebrates*, pp. 583–598. Verlag Dr. Friedrich Pfeil, München.
- Arratia, G. and Clouthier, R. 2002. Cheirolepidiform fish from the Devonian of Red Hill, Nevada. *Journal of Vertebrate Paleontology* 22: 33A.
- Ax, P. 1987. *The Phylogenetic System*. John Wiley and Sons, Chichester.
- Barton, D. G. and Wilson, M. V. H. 2005. Taphonomic variations in Eocene fish-bearing varves at Horsefly, British Columbia, reveal 10 000 years of environmental change. – *Canadian Journal of Earth Sciences* 42: 137–149.
- Behrensmeyer, A. K. 1978. Taphonomic and ecologic information from bone weathering. – *Paleobiology* 4: 150–162.
- Benton, M. J. 1993. *The Fossil Record 2*. Chapman & Hall, London.
- Benton, M. J. 2005. *Vertebrate Palaeontology*, 3rd edn. Blackwell Science Ltd, Oxford.
- Benton, M. J. and Donoghue, P. C. J. 2007. Paleontological evidence to date the tree of life. – *Molecular Biology and Evolution* 24: 26–53.
- Bird, N. C. and Mabee, P. M. 2002. Homology, individuality, and developmental morphology of the axial skeleton of the zebrafish. *Integrative and Comparative Biology* 42: 1197.
- Briggs, D. E. G. and Fortey, R. A. 2005. Wonderful strife: Systematics, stemgroups, and the phylogenetic signal of the Cambrian radiation. – *Paleobiology* 31: 94–112.
- Brown, J. H. 1999. Macroecology: Progress and prospect. – *Oikos* 87: 3–14.
- Budd, G. E. and Jensen, S. 2000. A critical reappraisal of the fossil record of the bilaterian phyla. – *Biological Reviews* 75: 253–295.
- Burness, G. P., Diamond, J. and Flannery, T. 2001. Dinosaurs, dragons, and dwarfs: The evolution of maximal body size. – *Proceedings of the Academy of Nat Sciences of the United States of America* 98: 14518–14523.
- Butler, M. A. and Losos, J. B. 1997. Testing for unequal amounts of evolution in a continuous character on different branches of a phylogenetic tree using linear and squared-change parsimony: An example using lesser antillean *Anolis* lizards. – *Evolution* 51: 1623–1635.
- Butler, V. L. 2004. Fish remains. In Hicks, B. A. (Ed.): *Marmes Rockshelter A Final Report on 11 000 Years of Cultural Use*, pp. 319–337. Washington State University Press, Pullman.
- Butler, V. L. and Schroeder, A. R. 1998. Do digestive processes leave diagnostic traces on fish bones? *Journal of Archaeological Science* 25: 957–971.
- Clouthier, R. 1996. The primitive actinistian *Miguashaia bureaui* Schultz (Sarcopterygii) In Schultze, H. P. and Clouthier, P. (Eds): *Devonian Fishes and Plants of Miguasha, Quebec, Canada*, pp. 227–247. Verlag Dr. Friedrich Pfeil, München.
- Clouthier, R. 1997. Morphologie et variations du toit crânien du Dipneuste *Scaumenacia curta* (Whiteaves; Sarcopterygii), du Dévonien supérieur du Québec. – *Geodiversitas* 19: 59–105.
- Clouthier, R. and Forey, P. L. 1991. Diversity of extinct and living actinistian fishes (Sarcopterygii). – *Environmental Biology of Fishes* 32: 59–74.
- Clouthier, R. and Ahlberg, P. E. 1996. Morphology, characters, and the interrelationships of basal sarcopterygians. In Stiassny, M. L., Parenti, L. R. and Johnson, G. D. (Eds): *Interrelationships of Fishes*, pp. 445–479. Academic Press, San Diego.
- Coates, M. I. 1998. Actinopterygians from the Namurian of Bearsden, Scotland, with comments on early actinopterygian neurocrania. – *Zoological Journal of the Linnean Society* 122: 27–59.
- Cobbett, A., Wilkinson, M. and Wills, M. A. 2007. Fossils impact as hard as living taxa in parsimony analyses of morphology. – *Systematic Biology* 56: 753–766.
- Conway Morris, S. 1993. The fossil record and the early evolution of the Metazoa. – *Nature* 361: 219–225.
- Cooper, R. A., Maxwell, P. A., Crampton, J. S., Beu, A. G., Jones, C. M. and Marshall, B. A. 2006. Completeness of the fossil record: Estimating losses due to small body size. – *Geology* 34: 241–244.
- Crampton, J. S., Beu, A. G., Cooper, R. A., Jones, C. M., Marshall, B. A. and Maxwell, P. A. 2003. Estimating the rock volume bias in paleobiodiversity studies. – *Science* 30: 358–360.
- Crimes, T. P. and Droser, M. L. 1992. Trace fossils and bioturbation: The other fossil record. – *Annual Review of Ecology and Systematics* 23: 339–360.
- Crisp, M. D. and Cook, L. G. 2005. Do early branching lineages signify ancestral traits. – *Trends in Ecology and Evolution* 20: 122–128.
- Dietze, K. 2000. A revision of paramblypterid and amplypterid actinopterygians from Upper Carboniferous–Lower Permian lacustrine deposits of central Europe. – *Palaeontology* 43: 927–966.
- Dong, X. P., Donoghue, P. C. J. and Repetski, J. E. 2005. Basal tissue structure in the earliest euconodonts: Testing hypotheses of developmental plasticity in euconodont phylogeny. – *Palaeontology* 48: 411–421.
- Donoghue, M. J., Doyle, J. A., Gauthier, J., Kluge, A. G. and Rowe, T. 1989. The importance of fossils in phylogeny reconstruction. – *Annual Review of Ecology and Systematics* 20: 431–460.

- Donoghue, P. C. J. and Sansom, I. J. 2002. Origin and early evolution of vertebrate skeletonization. – *Microscopy Research and Technique* **59**: 352–372.
- Finarelli, J. A. and Flynn, J. J. 2006. Ancestral state reconstruction of body size in the Caniformia (Carnivora, Mammalia): The effects of incorporating data from the fossil record. – *Systematic Biology* **55**: 301–313.
- Friedman, M. and Blom, H. 2006. A new actinopterygian from the Famennian of east Greenland and the interrelationships of Devonian ray-finned fishes. – *Journal of Paleontology* **80**: 1186–1204.
- Froese, R. and Binohlan, C. 2000. Empirical relationships to estimate asymptotic length, length at first maturity and length at maximum yield per recruit in fishes with simple methods to evaluate length frequency data. – *Journal of Fish Biology* **56**: 758–773.
- Froese, R. and Pauly, D. 2005. *Fishbase 2005: Concepts, Design and Data Sources*. ICLARM, Los Banos.
- Gardezi, T. and da Silva, J. 1999. Diversity in relation to body size in mammals: A comparative study. – *American Naturalist* **153**: 110–123.
- Gatesy, J. and O’Leary, M. A. 2001. Deciphering whale origins with molecules and fossils. *Trends in Ecology and Evolution* **16**: 658–658.
- Gauthier, J., Kluge, A. G. and Rowe, T. 1988. Amniote phylogeny and the importance of fossils. – *Cladistics* **4**: 105–209.
- Gess, R. W., Coates, M. I. and Rubidge, B. S. 2006. A lamprey from the Devonian period of South Africa. – *Nature* **443**: 981–984.
- Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M. and Charnov, E. L. 2001. Effects of size and temperature on metabolic rate. – *Science* **293**: 2248–2251.
- Gradstein, F. M., Ogg, J. G., Smith, A. G., Agterberg, F. P., Bleeker, W., Cooper, R. A. et al. 2004. *A Geologic Time Scale*. Cambridge University Press, Cambridge.
- Haldane, J. B. S. 1985. *On Being the Right Size and Other Essays*. Oxford University Press, Oxford.
- Hone, D. W. E. and Benton, M. J. 2005. The evolution of large size: How does Cope’s Rule work? *Trends in Ecology and Evolution* **20**: 4–6.
- Hurley, I. A., Mueller, R. L., Dunn, K. A., Schmidt, E. J., Friedman, M., Ho, R. K., Prince, V. E., Yang, Z. H., Thomas, M. G. and Coates, M. I. 2007. A new time-scale for ray-finned fish evolution. – *Proceedings of the Royal Society of B-Biological Sciences* **274**: 489–498.
- Hutchinson, G. E. and MacArthur, R. H. 1959. A theoretical ecological model of size distributions among species of animals. – *American Naturalist* **93**: 117–125.
- Janvier, P. 1996. *Early Vertebrates*. Oxford University Press, Oxford.
- Janvier, P. 2003. Vertebrate characters and the Cambrian vertebrates. – *Comptes Rendus Palevol* **2**: 523–531.
- Janvier, P., Desbiens, S., Willett, J. A. and Arsenaault, M. 2006. Lamprey-like gills in a gnathostome-related Devonian jawless vertebrate. – *Nature* **440**: 1183–1185.
- Janvier, P. and Lund, R. 1983. *Hardistiella montanensis* N. General et sp. (Petromyzontida) from the Lower Carboniferous of Montana, with remarks on the affinities of the lampreys. – *Journal of Vertebrate Paleontology* **2**: 407–413.
- Kidwell, S. M. and Flessa, K. W. 1995. The quality of the fossil record: Populations, species, and communities. – *Annual Review of Ecology and Systematics* **26**: 269–299.
- Knouft, J. H. and Page, L. M. 2003. The evolution of body size in extant groups of North American freshwater fishes: Speciation, size distributions, and Cope’s rule. – *American Naturalist* **161**: 413–421.
- Lane, A., Janis, C. M. and Sepkoski Jr., J. J. 2005. Estimating paleo-diversities: A test of the taxic and phylogenetic methods. – *Paleobiology* **31**: 21–34.
- Laurin, M. 2004. The evolution of body size, Cope’s rule and the origin of amniotes. – *Systematic Biology* **53**: 594–622.
- Leaper, R., Raffaelli, D., Emes, C. and Manly, B. 2001. Constraints on body-size distributions: An experimental test of the habitat architecture hypothesis. – *Journal of Animal Ecology* **70**: 248–259.
- Long, D. J. and Langer, M. R. 1995. Nassariid gastropods as destructive agents in preservation and fossilization of marine fishes. – *Cellular and Molecular Life Sciences* **51**: 85–89.
- Long, J. A. 1995. *The Rise of Fishes: 500 Million Years of Evolution*. Johns Hopkins University Press, Baltimore & London.
- Lund, R. 2000. The new actinopterygian order Guildayichthyiformes from the Lower Carboniferous of Montana (USA). – *Geodiversitas* **22**: 171–206.
- Lund, R. and Poplin, C. 2002. Cladistic analysis of the relationships of the Tarrasiids (Lower Carboniferous Actinopterygians). – *Journal of Vertebrate Paleontology* **22**: 480–486.
- Mabee, P. M. 2002. Evolution of fin modules in the axial skeleton of fishes. *Integrative and Comparative Biology* **42**: 1270.
- Mabee, P. M. 2000. Developmental data and phylogenetic systematics: Evolution of the vertebrate limb. – *American Zoologist* **40**: 789–800.
- Maddison, W. P. 1991. Squared-change parsimony reconstructions of ancestral states for continuous-valued characters on a phylogenetic tree. – *Systematic Zoology* **40**: 304–314.
- Maddison, W. P. and Maddison, D. R. 2005. *MACCLADE, Analysis of Phylogeny and Character Evolution*, Version 4.07. Sunderland Associates, Inc, Massachusetts.
- Maddison, W. P. and Maddison, D. R. 2006. *MESQUITE: A Modular System for Evolutionary Analysis*, Version 1.11. <http://mesquite-project.org>.
- Madin, J. S., Alroy, J., Aberhan, M., Fursich, F. T., Kiessling, W., Kosnik, M. A. and Wagner, P. J. 2006. Statistical independence of escalatory ecological trends in Phanerozoic marine invertebrates. – *Science* **312**: 897–900.
- Mallatt, J. and Chen, J. Y. 2003. Fossil sister group of craniates: Predicted and found. – *Journal of Morphology* **258**: 1–31.
- McGhee, G. R. 1996. *The Late Devonian Mass Extinction: The Frasnian/Famennian Crisis*. – Columbia University Press, New York.
- McNab, B. K. 2002. *Physiological Ecology of Vertebrates: A View from Energetics*. – Cornell University Press, Ithaca.
- Northcutt, R. G. 2005. The new head hypothesis revisited. – *Journal of Experimental Zoology Part B-Molecular and Developmental Evolution* **304B**: 274–297.
- Northwood, C. 2005. Early Triassic coprolites from Australia and their palaeobiological significance. – *Palaeontology* **48**: 49–68.
- O’Leary, M. A. 1999. Parsimony analysis of total evidence from extinct and extant taxa and the Cetacean-Artiodactyl question (Mammalia, Ungulata). – *Cladistics* **15**: 315–330.
- O’Leary, M. A. 2001. The phylogenetic position of cetaceans: Further combined data analyses, comparisons with the stratigraphic record and a discussion of character optimization. – *American Zoologist* **41**: 487–506.
- Pagel, M. 1999. Inferring the historical patterns of biological evolution. – *Nature* **401**: 877–884.
- Pagel, M., Venditti, C. and Meade, A. 2006. Large punctuational contribution of speciation to evolutionary divergence at the molecular level. – *Science* **314**: 119–121.
- Patterson, C. 1981. Significance of fossils in determining evolutionary relationships. – *Annual Review of Ecology and Systematics* **12**: 195–223.

- Peters, R. H. 1983. *The Ecological Implications of Body Size*. Cambridge University Press, Cambridge.
- Peters, S. E. and Foote, M. 2001. Biodiversity in the Phanerozoic: A reinterpretation. – *Paleobiology* 27: 583–601.
- Pollock, D. D., Zwickl, D. J., McGuire, J. A. and Hillis, D. M. 2002. Increased taxon sampling is advantageous for phylogenetic inference. – *Systematic Biology* 51: 664–671.
- Polly, P. D. 1998. Cope's rule. – *Science* 282: 50–51.
- Polly, P. D. 2001. Paleontology and the comparative method: Ancestral node reconstructions versus observed node values. – *American Naturalist* 157: 596–609.
- Prendini, L. 2001. Species or supraspecific taxa as terminals in cladistic analysis? Groundplans versus exemplars revisited. – *Systematic Biology* 50: 290–300.
- Prendini, L. and Wheeler, W. C. 2004. Assembling the scorpion tree of life: Phylogeny of extant scorpions based on morphology, molecules and exemplars. – *Cladistics* 20: 602–603.
- Santini, F. and Tyler, J. C. 2004. The importance of even highly incomplete fossil taxa in reconstructing the phylogenetic relationships of the Tetraodontiformes (Acanthomorpha: Pisces). – *Integrative and Comparative Biology* 44: 349–357.
- Schmidt-Nielsen, K. 1984. *Scaling Why is Animal Size so Important?* Cambridge University Press, Cambridge.
- Sepkoski, J. J. 2002. A compendium of fossil marine animal genera. In Jablonski, D. and Foote, M. (Eds): *Bulletins of American Paleontology*, pp. 1–560. Paleontological Research Institution, Ithaca.
- Shu, D. G. 2003. A paleontological perspective of vertebrate origin. – *Chinese Science Bulletin* 48: 725–735.
- Shu, D. G., Morris, S. C., Han, J., Zhang, Z. F., Yasui, K., Janvier, P., Chen, L., Zhang, X. L., Liu, J. N., Li, Y. and Li, H. Q. 2003. Head and backbone of the Early Cambrian vertebrate *Haikouichthys*. – *Nature* 421: 526–529.
- Simpson, G. G. 1952. How many species? *Evolution* 6: 342.
- Smith, F. A., Brown, J. H., Haskell, J. P., Lyons, S. K., Alroy, J., Charnov, E. L. et al. 2004. Similarity of mammalian body size across the taxonomic hierarchy and across space and time. – *American Naturalist* 163: 672–691.
- Stanley, S. M. 1973. An explanation for Cope's rule. – *Evolution* 27: 1–26.
- Stanley, S. M. 1998. *Macroevolution. Pattern and Process*. The Johns Hopkins University Press, Baltimore.
- Strathdee, A. T. and Bale, J. S. 1998. Life on the edge: Insect ecology in Arctic environments. – *Annual Review of Entomology* 43: 85–106.
- Swofford, D. L. and Maddison, W. P. 1987. Reconstructing ancestral character states under Wagner parsimony. – *Mathematical Biosciences* 87: 199–229.
- Twitchett, R. J., Feinberg, J. M., O'Connor, D. D., Alvarez, W. and McCollum, L. B. 2005. Early Triassic ophiuroids: Their paleoecology, taphonomy, and distribution. – *Palaio* 20: 213–223.
- Underwood, C. J. 2006. Diversification of the Neoselachii (Chondrichthyes) during the Jurassic and Cretaceous. – *Paleobiology* 32: 215–235.
- Valentine, J. W., Jablonski, D., Kidwell, S. and Roy, K. 2006. Assessing the fidelity of the fossil record by using marine bivalves. – *Proceedings of the National Academy of Sciences of the United States of America* 103: 6599–6604.
- Vermeij, G. J. 1993. *Evolution and Escalation: An Ecological History of Life*. Princeton University Press, Princeton.
- Vermeij, G. J. 1994. The evolutionary interaction among species: Selection, escalation, and coevolution. – *Annual Review of Ecology and Systematics* 25: 219–236.
- Wake, D. B. 1991. Homoplasy: the result of natural selection or evidence of design limitations. – *American Naturalist* 138: 543–567.
- Webby, B., Paris, D., Droser, M. L. and Percival, I. G. 2004. *The Great Ordovician Biodiversification Event*. Columbia University Press, New York.
- Webster, A. J., Gittleman, J. L. and Purvis, A. 2004. The life history legacy of evolutionary body size change in carnivores. – *Journal of Evolutionary Biology* 17: 396–407.
- Wickstrom, L. M. and Donoghue, P. C. J. 2005. Cladograms, phylogenies and the veracity of the conodont fossil record. *Conodont Biology and Phylogeny: Interpreting the Fossil Record*, pp. 185–218. The Palaeontological Association, Cambridge, UK.
- Wiens, J. J. 2006. Missing data and the design of phylogenetic analyses. – *Journal of Biomedical Informatics* 39: 34–42.
- Wilkinson, M. 2003. Missing entries and multiple trees: Instability, relationships, and support in parsimony analysis. – *Journal of Vertebrate Paleontology* 23: 311–323.
- Wilson, M. V. H. 1992. Importance for phylogeny of single and multiple stem-group fossil species, with examples from fresh-water fishes. – *Systematic Biology* 41: 462–470.

Appendix 1. The fossil fish dataset. Size and stratigraphic data for 465 fish species, including 425 species known only as fossils and 40 extant taxa for clades lacking fossils with reliable size information. Data for 15 extant taxa presented as averages from a total of 160 species as follows: 7 Myxiniiformes (69 spp.); 2 Petromyzontiformes (40 spp.); *Protopterus* (6 spp.); *Polypterus* (17 spp.); *Acipenser* (18 spp.); *Scaphyrinchus* (2 sp.); *Pseudoscaphyrinchus* (3 spp.); *Clupea* (5 spp.). Minimum geological ages (Ma) from stratigraphic data to Series or Epoch from (Benton 1993) (Benton 2005). Taxa arranged according to conventional phylogenetic sequence. TL, maximum known total length (cm).

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference	
Cephalochordata	† <i>Cathaymyrus diadexus</i>	LC	530	5.0	(1)	
	† <i>Pikaea gracilens</i>	MC	513	3.0	(1)	
	† <i>Paleobranchiostoma hamatotergum</i>	MC	513	4.0	(2)	
Yunnanozoa	<i>Branchiostoma</i> 16 spp.	R	0	4.0	(3)	
	† <i>Yunnanozoon lividum</i>	LC	530	3.0	(4)	
Hyperotreti	† <i>Haikouella lanceolatum</i>	LC	530	3.0	(5) (6)	
	† <i>Gilpichthys greeniei</i>	Pn	318	8.0	P. Janvier, pers. comm.	
	† <i>Myxinkela siroka</i>	Pn	318	7.0	P. Janvier, pers. comm.	
	† <i>Myxineides gononorum</i>	Pn	318	15.0	P. Janvier, pers. comm.	
	<i>Eptatretus</i>	R	0	53.0	(3)	
	<i>Paramyxine</i>	R	0	43.0	(3)	
	<i>Quadratus</i>	R	0	28.0	(3)	
	<i>Myxine</i>	R	0	55.0	(3)	
	<i>Nemamyxine</i>	R	0	51.0	(3)	
	<i>Neomyxine</i>	R	0	41.0	(3)	
Myllokunmingiida	† <i>Myllokunmingia fengjiaoa</i>	LC	530	2.8	(7)	
	† <i>Zhongjianichthys rostratus</i>	LC	530	2.5	(7)	
Hyperoartia	† <i>Haikouichthys ercaicunensis</i>	LC	530	2.5	(8)	
	† <i>Endeiolepis aneri</i>	UD	385	10.0	(9)	
	† <i>Legendrelepis parenti</i>	UD	385	10.0	(10)	
	† <i>Jamoytius kerkwoodi</i>	LS	444	19.0	(10)	
	† <i>Euphanerops longaevus</i>	UD	385	10.0	(9)	
	† <i>Hardistiella montanensis</i>	M	360	10.0	(11)	
	† <i>Mayomyzon pieckoensis</i>	Pn	318	6.0	(10)	
	† <i>Pipiscius zangerli</i>	M	360	6.2	P. Janvier, pers. comm.	
	Geotridae	R	0	36.0	(3)	
	Petromyzontidae	R	0	30.0	(3)	
	Pteraspodomorphi	† <i>Andinaspis suarezorum</i>	LO	488	15.0	(12)
		† <i>Anatolepis sp.</i>	UC	501	8.0	(13)
		† <i>Arandaspis prionotolepis</i>	LO	488	14.0	(14)
† <i>Porophoraspis crenulata</i>		LO	488	30.0	P. Janvier, pers. comm.	
† <i>Sacabambaspis janvieri</i>		UO	461	35.0	(10)	
† <i>Astraspis splendens</i>		UO	461	40.0	(15)	
† <i>Lepidaspis serrata</i>		D	416	25.0	(10)	
† <i>Empedaspis inermis</i>		LD	416	7.0	(10)	
† <i>Athenaegis chattertoni</i>		LS	444	6.0	(10)	
† <i>Irregulareaspis hoeli</i>		LD	416	12.0	(10)	
† <i>Pionaspis amplissima</i>		LD	416	20.0	(10) (9)	
† <i>Torpedaspis elongata</i>		LD	416	25.0	(10)	
† <i>Vernonaspis sp.</i>		US	423	10.0	(10)	
† <i>Anglaspis insignis</i>		LD	416	5.0	(10)	
† <i>Canadapteraspis allocostomata</i>		LD	416	12.0	(10) (16)	
† <i>Trygonaspis sp.</i>		LD	416	12.0	(10)	
† <i>Tolypelepis undulata</i>		US	423	25.0	(17)	
† <i>Cardipeltis bryanti</i>		LD	416	19.0	(10)	
† <i>Drepanaspis gemuendensis</i>		LD	416	53.0	(10)	
† <i>Drepanaspis sp.</i>		LD	416	100.0	(10)	
† <i>Doryaspis nathorsti</i>	LD	416	20.0	(10)		
† <i>Larnovaspis goujeti</i>	LD	416	20.0	(10)		
† <i>Protaspis transversa</i>	US-LD	423	30.0	(10)		
† <i>Pteraspis rostrata</i>	LD	416	20.0	(10)		
† <i>Rhinopteraspis sp.</i>	LD	416	30.0	(10)		
† <i>Psammolepis paradoxa</i>	UD	385	70.0	(9)		

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference	
Thelodonti	† <i>Loganellia scotica</i>	LO	488	40.0	(18, 19)	
	† <i>Phlebolepis elegans</i>	US	423	9.0	(10)	
	† <i>Shiella taiti</i>	MS	428	13.3	(20)	
	† <i>Loganellia scotia</i>	US	423	15.0	(10)	
	† <i>Lanarkia horrida</i>	US	423	10.0	(10)	
	† <i>Thelodus scotius</i>	US	423	15.0	(10)	
	† <i>Archipelepis turbinata</i>	LS	444	6.0	(19)	
	† <i>Turinia pagei</i>	LD	416	40.0	(10)	
Anaspida	† <i>Birkenia elegans</i>	US	423	10.0	(10)	
	† <i>Lasanius problematicus</i>	US	423	8.0	(10)	
	† <i>Pharyngolepis oblongus</i>	US	423	20.0	(10)	
	† <i>Pterygolepis nitidus</i>	US	423	10.0	(10)	
	† <i>Rhyncholepis parvulus</i>	LD	416	7.0	(10)	
Galeaspidi	† Birkeniidae unnamed	MS	428	15.0	(10)	
	† <i>Hanyangaspis guodinshanensis</i>	MS	428	30.0	P. Janvier, pers. comm.	
	† <i>Xiushuiaspis</i> sp.	LS	444	10.0	(9)	
	† <i>Sinoszechuanaspis longicornis</i>	LD	416	8.0	P. Janvier, pers. comm.	
	† <i>Polybranchiaspis</i> sp.	LD	416	8.5	(9)	
	† <i>Bannhuanaspis</i> sp.	LD	416	20.0	(9)	
	† <i>Huananaspis guodinshanensis</i>	MS	428	25.0	P. Janvier, pers. comm.	
Pituriaspida	† <i>Sanqiaspis</i> sp.	LD	416	12.0	(9)	
	† <i>Pituriaspis doylei</i>	LD	416	25.0	(9)	
Osteostraci	† <i>Atelaspis tessellata</i>	US	423	20.0	(9)	
	† <i>Hemicyclaspis murchisoni</i>	US	423	18.0	(10)	
	† <i>Hirella gracilis</i>	US	423	7.0	(10)	
	† <i>Witaaspis</i> sp.	US	423	12.0	(9)	
	† <i>Dartmuthia</i> sp.	US	423	18.0	(9)	
	† <i>Tyriaspis whitei</i>	US	423	10.0	(10)	
	† <i>Atelaspis robustus</i>	LD	416	15.0	(10)	
	† <i>Boreaspis puella</i>	LD	416	8.0	(10)	
	† <i>Cephalaspis powerei</i>	LD	416	20.0	(10)	
	† <i>Zenaspis pagei</i>	LD	416	25.0	(9)	
	† <i>Parameteoraspis</i> sp.	LD	416	100.0	(16)	
	† <i>Procephalaspis</i> sp.	US	423	49.0	(9)	
	† <i>Norselaspis glacialis</i>	LD	416	8.0	(9)	
	† <i>Escuminaspis laticeps</i>	UD	385	25.0	(9)	
	† <i>Alaspis microtuberculata</i>	UD	385	31.0	(10)	
	Furcacaudiformes	† <i>Furcacauda heintzae</i>	LD	416	9.0	UALVP 32958
		† <i>Sphenonectris turnerae</i>	LD	416	12.0	UALVP 42212
	Placodermi	† <i>Stensioella heintzi</i>	LD	416	26.0	(10)
		† <i>Pseudopetalichthys problematicus</i>	LD	416	18.0	(15)
		† <i>Antarctaspis</i> sp.	MD	398	30.0	(9)
† <i>Austrophyllolepis</i> sp.		MD	398	20.0	(9)	
† <i>Wuttagoonaspis fletsheri</i>		MD	398	30.0	(9)	
† <i>Sigaspis lepidophora</i>		LD	416	20.0	(9)	
† <i>Dicksonosteus</i> sp.		LD-MD	416	20.0	(9)	
† <i>Gemuendenaspis</i> sp.		LD	416	30.0	(9)	
† <i>Tiaraspis</i> sp.		LD	416	24.0	(9)	
† <i>Holonema rugosum</i>		MD-UD	398	70.0	(9)	
† <i>Homosteus milleri</i>		MD	398	375.0	(9)	
† <i>Antineosteus</i> sp.		LD	416	40.0	(9)	
† <i>Oxyosteus</i> sp.		UD	385	100.0	(10)	
† <i>Cocosteus decipiens</i>		MD	398	60.0	(9)	
† <i>Millerosteus minor</i>		MD	398	10.0	(10)	
† <i>Watsonosteus fletti</i>		MD	398	60.0	(10)	
† <i>Plourdosteus canadensis</i>		UD	385	100.0	(9)	
† <i>Eastmanosteus</i>	MD-UD	398	300.0	(10)		
† <i>Dunkleosteus terrelli</i>	UD	385	800.0	(10)		
† <i>Brachyosteus dietrichi</i>	UD	385	25.0	(10)		

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference
	† <i>Pholidosteus friedelti</i>	UD	385	85.0	(10)
	† <i>Phyllolepis</i> sp.	UD	385	40.0	(10)
	† <i>Lunaspis brolii</i>	LD	416	30.0	(10)
	† <i>Macropetalichthys sullivanii</i>	MD	398	80.0	(10)
	† <i>Rhamphodopsis threiplandi</i>	MD	398	7.0	(15)
	† <i>Ctenurella gladbachensis</i>	UD	385	18.0	(10)
	† <i>Rhamphodopsis trispinatus</i>	MD	398	12.0	(10)
	† <i>Yunnanolepis spinulosa</i>	LD	416	5.0	(21)
	† <i>Chuchinolepis spinulosa</i>	LD	416	7.5	(9)
	† <i>Sinolepis</i> sp.	LD	416	9.0	(9)
	† <i>Bothrolepis canadensis</i>	MD-UD	398	50.0	(10)
	† <i>Bothriolepis yeungae</i>	UD	385	50.0	(22)
	† <i>Diplognathus mirabilis</i>	UD	385	45.0	(10)
	† <i>Pterichthyodes milleri</i>	LD-MD	416	30.0	(10)
	† <i>Asterolepis maxima</i>	MD-UD	398	70.0	(10)
	† <i>Microbrachius dicki</i>	MD	398	6.0	(10)
	† <i>Remigolepis walkeri</i>	UD	385	40.0	(10)
	† <i>Gemuendia stuertzi</i>	LD	416	100.0	(10)
	† <i>Jagorina pandora</i>	UD	385	19.0	(10)
Chondrichthyes	† <i>Obtusacanthus corroconis</i>	LD	416	12.0	(23)
	† <i>Lupopsyroides macracanthus</i>	LD	416	4.0	(23)
	† <i>Diademodus hydei</i>	UD	385	39.0	(10)
	† <i>Cladoselache clarki</i>	UD-P	385	300.0	(24)
	† <i>Helicoprion besonovi</i>	LP	299	100.0	(10)
	† Eugeneodontidae unnamed	Tr	251	80.0	(10)
	† <i>Caseodus eatoni</i>	P	299	95.0	(9)
	† <i>Fadenia crenulata</i>	Pn	318	133.3	(9)
	† <i>Heteropetalus elegantulus</i>	M	360	12.0	(10)
	† <i>Belantsea montana</i>	M	360	27.5	(10, 24)
	† <i>Janassa bituminosa</i>	M-LP	360	54.0	(10)
	† <i>Stethacanthus tumidus</i>	UD-Pn	385	150.0	(24)
	† <i>Orestiacanthus fergusi</i>	Pn	318	28.0	(25)
	† <i>Damocles serratus</i>	M	360	20.0	(10)
	† <i>Falcatus falcatus</i>	M	360	15.0	(10, 24)
	† <i>Symorium reniforme</i>	M-Pn	360	300.0	(10)
	† <i>Cobelodus aculeata</i>	M	360	158.3	(9)
	† <i>Helodus simplex</i>	M	360	45.0	(15)
	† <i>Chondrenchelys problematica</i>	M	360	20.0	(15)
	† <i>Harpagofututor volsellorhinus</i>	M	360	10.0	(10)
	† <i>Deltoptychius</i> sp.	M-Pn	360	50.0	(10)
	† <i>Menaspis armata</i>	UP	260	16.0	(10)
	† Cochliodontofomes unnamed 1	M	360	14.0	(10)
	† Cochliodontofomes unnamed 2	M	360	11.0	(10)
	† Cochliodontofomes unnamed 3	M	360	10.0	(10)
	† Cochliodontofomes unnamed 4	M	360	9.0	(10)
	† <i>Echinochimaera meltoni</i>	M	360	8.0	(10)
	† <i>Delphyodontos dacrifformes</i>	M	360	11.0	(10)
	† <i>Acanthorhina jaekeli</i>	LJ	200	50.0	(10)
	† <i>Ischyodus quenstedti</i>	MJ-PC	161	142.0	(10)
	† <i>Ischyodus avitus</i>	UJ	161	84.0	(26)
	<i>Chimaera monstrosa</i>	R	0	150.0	(3)
	<i>Hydrolagus affinis</i>	R	0	130.0	(3)
	<i>Rhinochimaera pacifica</i>	R	0	130.0	(3)
	† Iniopterygiiformes unnamed 1	M	360	10.0	(10)
	† Iniopterygiiformes unnamed 2	M	360	10.0	(10)
	† <i>Iniopteryx rushloui</i>	Pn	318	24.0	(10)
	† <i>Iniopera richardsoni</i>	Pn	318	24.0	(10)
	† <i>Sibirhynchus denisoni</i>	Pn	318	20.0	(10)
	† <i>Doliodus problematicus</i>	LD	416	75.0	(27)

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference
	† Elasmobranchii <i>unnamed</i>	M	360	18.0	(10)
	† <i>Antarctilamna prisca</i>	MD	398	60.0	(9, 28)
	† <i>Expleuracanthus gaudryi</i>	Pn	318	58.0	(10)
	† <i>Orthacanthus senckenbegianus</i>	LP	299	300.0	(26)
	† <i>Triodus sesselis</i>	LP	299	50.0	(10)
	† <i>Xenacanthus meisenheimensis</i>	LP	299	75.0	(10, 24)
	† <i>Ctenacanthus costellatus</i>	UD	385	150.0	(15) (24)
	† <i>Goodrichichthys sp.</i>	M	360	250.0	(15)
	† <i>Onychoselache traquairi</i>	M	360	24.0	(9)
	† <i>Hamiltonichthys mapesi</i>	Pn	318	28.0	(10)
	† <i>Hybodus hauffianis</i>	UP-UK	260	260.0	(10)
	† <i>Wodnika striaula</i>	UP	260	48.0	(10)
	† <i>Acronemus tuberculatus</i>	LTr-UK	251	29.0	(10)
	† <i>Heterodontus falcifer</i>	UJ-R	161	28.0	(10)
	† <i>Paracestracion zitteli</i>	UJ	161	15.0	(10)
	† <i>Notidanoides muensteri</i>	UJ	161	300.0	(29)
	† <i>Hexanchus gracilis</i>	UK	86	29.0	(10)
	† <i>Chlamydoselachus thompsni</i>	UK	100	200.0	(29)
	† <i>Chlamydoselachus lawleyi</i>	PL	5	200.0	(29)
	<i>Chlamydoselachus anguineus</i>	R	0	200.0	(3)
	† Orodontiformes <i>unnamed</i>	M	360	300.0	(10)
	† <i>Macrourogaleus hassei</i>	UJ	161	12.0	(10)
	† <i>Paleoscyllium formosum</i>	UJ-UK	161	60.0	(10)
	† <i>Scyliorhinus elongatus</i>	UK	100	23.0	(10)
	† Scyliorhinidae <i>unammed</i>	UK	100	87.0	(10)
	† <i>Paratriakis curtirostris</i>	UK	100	29.0	(10)
	† <i>Carcharodon (=Carcharocles?) megalodon</i>	ME-PI	49	1200.0	(10)
	† <i>Eogaleus bolcensis</i>	ME	49	110.0	(10)
	† <i>Galeorhinus cuvieri</i>	ME	49	78.0	(10)
	† <i>Squalicorax falcatus</i>	UK-PC	100	188.0	(10)
	† <i>Scapanorhynchus lewisii</i>	UK	100	65.0	(10)
	† <i>Orectolobus jurrasicus</i>	UJ	161	30.0	(10)
	† <i>Paleocarcharius stromeri</i>	UJ	161	86.0	(10)
	† <i>Phorcynis catulina</i>	UJ	161	40.0	(10)
	† <i>Mesiteia emilliae</i>	UK	100	27.0	(10)
	† <i>Aellopus bugesiacus</i>	UJ	161	110.0	(10)
	† <i>Asterodemus platypterus</i>	UJ	161	46.0	(10)
	† <i>Belemnobatis sismondæ</i>	UJ	161	40.0	(10)
	† <i>Cyclobatis major</i>	LK	146	13.0	(10)
	† <i>Pararaja expansa</i>	UK	100	23.0	(10)
	† <i>Rhinobatos hakelensis</i>	UK	100	28.0	(10)
	† <i>Rhombopterygia rajoides</i>	UK	100	42.0	(10)
	† <i>Micropristis solonis</i>	UK	100	53.0	(10)
	† <i>Sclerorhynchus atavus</i>	UK	100	100.0	(10)
	† <i>Heliobatis radiens</i>	EC	57	40.0	(10)
	† <i>Zapteryx bichuti</i>	EC	57	47.0	(10)
	† <i>Trygon muricata</i>	ME	49	73.0	(10)
	† <i>Promylobatis gazoae</i>	ME	49	45.0	(10)
	† <i>Platyrhina egertoni</i>	ME	49	55.0	(10)
	† <i>Trigonorhina deznii</i>	ME	49	79.0	(10)
	† <i>Urolophus crassicaudatus</i>	ME	49	80.0	(10)
	† <i>Protospinax annectens</i>	UJ	161	146.0	(10)
	† <i>Centrophoroides latidens</i>	UK	100	37.0	(10)
	† <i>Pseudothina alifera</i>	UJ	161	96.0	(10)
	† <i>Torpedo sp.</i>	ME	49	38.0	(10)
	† <i>Narcine molini</i>	LE	41	90.0	(10)
Acanthodii	† <i>Lupopsyrus pygmaeus</i>	LD	416	3.3	(30)
	† <i>Climatius reticulatus</i>	US-LD	423	14.0	(10, 17)
	† <i>Mesacanthus mitchelli</i>	LD-MD	416	15.0	(10, 17)

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference
	† <i>Cheiracanthus latus</i>	UD	385	30.0	(10)
	† <i>Homalacanthus concinnus</i>	UD-M	385	15.0	(10)
	† <i>Triazeugacanthus affinis</i>	UD	385	10.0	(10)
	† <i>Acanthodes bronni</i>	Pn-LP	318	50.0	(10, 17)
	† <i>Utahacanthus guntheri</i>	Pn	318	10.0	(10)
	† <i>Traquairichthys pygmaeus</i>	Pn-LP	318	10.0	(10, 17)
	† <i>Paucicanthus vanelsti</i>	LD	416	4.0	(31)
	† <i>Poracanthodes menneri</i>	US-LD	423	3.3	(30)
	† <i>Euthacanthus macnicolli</i>	LD	416	14.0	(10)
	† <i>Parexus falcatus</i>	LD	416	14.0	(10)
	† <i>Vernicomacanthus uncinatus</i>	LD	416	15.0	(10)
	† <i>Diplacanthus striatus</i>	MD-UD	398	20.0	(26)
	† <i>Rhadinacanthus longispinatus</i>	MD	398	16.0	(10)
	† <i>Ischnicanthus gracilis</i>	LD	416	35.0	(10)
Sarcopterygii	† <i>Psarolepis romeri</i>	US-LD	423	10.0	(32)
	† <i>Diabolepis speratus</i>	LD	416	30.0	(33)
	† <i>Orychodus sigmoides</i>	LD	416	200.0	(9)
	† <i>Strunius walteri</i>	MD	398	10.0	(15)
	† <i>Miguashaia bureaui</i>	MD	398	50.0	(9) (34)
	† <i>Diplurus macropterus</i>	LP	299	25.0	(10)
	† <i>Diplurus newarki</i>	UD	385	25.0	(15)
	† <i>Lochmocerurus aciculiodontus</i>	M	360	11.0	(10)
	† <i>Allenypterus montanus</i>	M	360	14.0	(10)
	† <i>Coelacanthus granulatus</i>	P	299	56.5	(15)
	† <i>Whiteia woodwardi</i>	LTr	251	75.0	(35)
	† <i>Rhabdoderma elegans</i>	UTr	228	15.0	(26)
	† <i>Axelrodichthys araripensis</i>	LK	146	70.0	(26)
	† <i>Macropoma lewesiensis</i>	UK	85	60.0	(35–37)
	<i>Latimeria</i> spp.	R	0	180.0	(3)
	† <i>Uranolophus wyomingensis</i>	LD	416	100.0	(38)
	† <i>Holoptychius quebecensis</i>	UD	385	43.0	(26)
	† <i>Porolepis elongata</i>	LD	416	100.0	(9, 24)
	† <i>Porolepis brevis</i>	MD	398	20.0	(10)
	† <i>Laccognathus panderi</i>	UD	385	100.0	(9)
	† <i>Youngolepis preacursor</i>	LD	416	30.0	(33)
	† <i>Holodipterus longi</i>	UD	385	45.0	(9)
	† <i>Dipterus valenciennesi</i>	MD	398	22.0	(15)
	† <i>Griphognathus whitei</i>	UD	385	70.0	(9)
	† <i>Scaumenacia curta</i>	UD	385	60.0	(26, 39)
	† <i>Phaneropleuron andersoni</i>	UD	385	20.0	(40)
	† <i>Ptychoceratodus philippi</i>	LTr	251	60.0	(41)
	<i>Neoceratodus forsteri</i>	UK	146	170.0	(3)
	† <i>Palaedaphus insignis</i>	UD	385	200	(28)
	† <i>Neoceratodus tuberculatus</i>	UK	100	200	(42)
	† <i>Rhinodipterus ulrichi</i>	UD	385	28	(43)
	† <i>Protopterus protopteroideus</i>	LK	146	70.0	(42)
	<i>Protopterus</i> spp.	EC	56	93.3	(3)
	<i>Lepidosiren paradoxa</i>	MC	23	125.0	(3)
	† <i>Strepsodus ancylonamensis</i>	M	360	30.0	(9)
	† <i>Osteolepis macrolepidotus</i>	MD	398	21.0	(15)
	† <i>Eusthenopteron foordi</i>	UD	385	25.0	(26)
	† <i>Elpistostege watsoni</i>	UD	385	50.0	(9)
	† <i>Panderichthys rhombolepis</i>	UD	385	145.0	(44)
	† <i>Sauripteris taylori</i>	UD	385	200.0	(26)
	† <i>Acanthostega gunnari</i>	UD	385	63.0	(26)
	† <i>Ichthyostega</i> sp.	UD	385	100.0	(45)
Actinopterygii	† <i>Andreolepis hedei</i>	US	423	25.0	(9, 46)
	† <i>Dialipina salgueiroensis</i>	LD	416	25.0	(47)
	† <i>Cheirolepis canadensis</i>	MD	398	30.0	(48, 49)

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference
	† <i>Cheirolepis tralii</i>	MD	398	25.0	(10)
	† <i>Cheirolepis schultzi</i>	MD	398	10.0	(48)
	† <i>Stegotrachelys finlayi</i>	MD	398	9.0	(10)
	† <i>Moythomasia nitida</i>	MD	398	10.0	(10, 14, 50)
	† <i>Howqualepis rostridens</i>	UD	385	95.0	(28, 50)
	† <i>Mimia paravertebrata</i>	UD	385	20.0	(9, 28, 50)
	† <i>Mentzichthys walsjhi</i>	UD	374	9.0	(10)
	† <i>Sundayichthys elegantulus</i>	UD	374	25.0	(10)
	† <i>Guildayichthys carnegie</i>	M	360	8	(51)
	† <i>Aetheretmon valenticum</i>	M	360	9.0	(10)
	† <i>Rhadinichthys carinatus</i>	M	360	11.0	(10)
	† <i>Canobius ramsayi</i>	M	360	6.0	(10)
	† <i>Coruboniscus budensis</i>	M	360	3.0	(10)
	† <i>Benedenus deneensis</i>	M	360	22.0	(10)
	† <i>Holurus parki</i>	M	360	12.0	(10)
	† <i>Nematoptychius greenocki</i>	M	360	45.0	(10)
	† <i>Drydenius insignis</i>	M	360	10.0	(10)
	† <i>Gonatodus punctatus</i>	M	360	17.0	(10)
	† <i>Acrolepis ortholepis</i>	M	360	30.0	(10)
	† <i>Acrolepis sedgwickii</i>	M	360	60.0	(52)
	† <i>Paramblypterus gelberti</i>	P	299	25.0	(53)
	† <i>Ganolepis gracilis</i>	P	299	7.0	(10)
	† <i>Acrolophis stensioei</i>	P	299	65.0	(10)
	† <i>Pygopterus humboldti</i>	UP	260	60.0	(52)
	† <i>Elonichthys punctatus</i>	LP	260	8.0	(10)
	† <i>Reticulepis exsculpta</i>	UP	260	57.0	(10)
	† <i>Bobasatriana canadensis</i>	Tr	251	67.0	(26)
	† <i>Apatolepis australis</i>	MTr	228	14.0	(10)
	† <i>Mesembroniscus longisquamosus</i>	MTr	228	8.0	(10)
	† <i>Polypterus dageti</i>	UK	98.6	54.1	(54)
	† <i>Mesopoma politum</i>	M	360	7.0	(10)
	† <i>Mesopoma planti</i>	Pn	318	7.0	(55)
	† <i>Dorypterus hoffmanni</i>	P	299	12.0	(52)
	† <i>Paleoniscum freislebeni</i>	UP	260	19.0	(26)
	† <i>Birgeria groenlandica</i>	LTr	251	84.0	(56)
	† <i>Saurichthys seefeldensis</i>	MTr	228	180.0	(9)
	† <i>Boreosomus gillioti</i>	LTr	251	13.0	(10)
	† <i>Perleidus madagascarensis</i>	LTr	251	12.0	(26)
	† <i>Redfieldius gracilis</i>	UTr	228	19.0	(26)
	† <i>Chondrosteus hindenburgi</i>	LJ	200	300.0	(10)
	† <i>Dapedium pholidotum</i>	LJ	200	26.0	(26)
	† <i>Peipiaosteus pani</i>	UJ	161	60.0	(56)
	† <i>Protopsephurus liui</i>	UJ	161	23.3	(52)
	† <i>Protoscaphirhynchus squamosus</i>	LK	146	56.0	(10)
	† <i>Polyodon tuberculata</i>	EC	55	260.5	(3)
	† <i>Crossopholis magnicaudata</i>	EC	55	39.0	(10)
	<i>Huso</i> spp.	R	0	530.0	(3)
	<i>Acipenser sinensis</i>	R	0	130.0	(3)
	<i>Scaphirhynchus</i> spp.	R	0	140.0	(3)
	<i>Pseudoscaphirhynchus</i>	R	0	56.0	(3)
	<i>Psephurus gladius</i>	R	0	300.0	(3)
	† <i>Paralepidotus ornatus</i>	UTr	228	30.0	(57)
	† <i>Semionotus agassizii</i>	UTr	228	29.0	(26)
	† <i>Pachchormus eoconus</i>	LJ	200	94.0	(26)
	† <i>Caturus velifer</i>	UJ	161	57.0	(26)
	† <i>Strobilodus giganteus</i>	UJ	161	177.0	(10)
	† <i>Amiopsis dolloi</i>	LK	146	14.0	(10)
	† <i>Calamopleurus cylindricus</i>	LK	146	91.0	(26)
	† <i>Lepisosteus simplex</i>	LK	146	73.0	(26)

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference
	† <i>Obaichthys decoratus</i>	LK	146	65.0	(26)
	† <i>Teoichthys kallistos</i>	LK	146	25.0	(26)
	† <i>Neoproscinetes penalvi</i>	LK	146	30.0	(26)
	<i>Amia calva</i>	EC	50	109.0	(3)
	† <i>Amia scutata</i>	EC	50	70.0	(58)
	† <i>Pholidophorus bechei</i>	LJ	200	20.0	(26)
	† <i>Parapholidophorus caffii</i>	UTr	228	8.0	(10)
	† <i>Pholidophorus macrocephalus</i>	LJ	200	33.0	(10)
	† <i>Aspidorhynchus acutirostris</i>	UJ	161	57.0	(26)
	† <i>Vinctifer comptoni</i>	LK	146	75.0	(26)
	† <i>Longileptolepis weidenothi</i>	LJ	183	37.4	(49)
	† <i>Humbertia operta</i>	UK	100	10.0	(10)
	† <i>Leptolepides haertesi</i>	UJ	150	5.4	(59)
	† <i>Leptolepides spratiformis</i>	UJ	150	9.1	(59)
	† <i>Longileptolepis wiedenrothi</i>	UJ	161	36.0	(49)
	† <i>Orthogonikleithrus hoelli</i>	UJ	161	4.5	(59)
	† <i>Orthogonikleithrus leichi</i>	UJ	161	10.1	(59)
	† <i>Cavenderichthys talbragarensis</i>	LJ	183	12.0	(10)
	† <i>Cladocyclus gardneri</i>	LK	146	111.0	(26)
	† <i>Gillicus arcuatus</i>	LK	146	157.0	(10)
	† <i>Ichthyodectes ctenodon</i>	LK	146	220.0	(10)
	† <i>Pachythrissops propterus</i>	UJ	150	38.0	(10)
	† <i>Thrissops formosus</i>	UJ	150	28.0	(10)
	† <i>Antarctithrissops australis</i>	UJ	150	30.0	(60)
	† <i>Xiphactinus audax</i>	UK	100	430.0	(10)
	† <i>Eohiodon falcatus</i>	EC	50	17.0	(10)
	<i>Hiodon tergisus</i>	EC	50	50.0	(59)
	† <i>Lycoptera davidi</i>	LK	146	8.0	(10)
	<i>Arapiama gigas</i>	MM	15	450.0	(61)
	<i>Heterotis niloticus</i>	R	0	100.0	(3)
	<i>Osteoglossum bicirrhosum</i>	R	0	120.0	(3)
	<i>Pantodon buchholzi</i>	R	0	10.0	(3)
	† <i>Phareodus testis</i>	EC	54	31.0	(10)
	† <i>Singida jacksonoides</i>	EC	54	18.0	(62)
	† <i>Chauliopareion mahengeense</i>	EC	45	9.0	(62)
	† <i>Lebonichthys lewisi</i>	UK	100	30.0	(10)
	† <i>Mylomyrus frangens</i>	EC	54	31.0	(10)
	† <i>Notelops brama</i>	LK	146	73.0	(10)
	† <i>Rhacolepis buccalis</i>	LK	146	14.0	(10)
	† <i>Spaniodon elongatus</i>	UK	100	18.0	(10)
	† <i>Brannerion sp.</i>	LK	146	45.0	(26)
	† <i>Elops sp.</i>	UJ	150	20.0	(59)
	† <i>Eomyrophis latispinus</i>	EC	57	24.0	(26)
	† <i>Paraelops cearensis</i>	LK	146	70.0	(10)
	† <i>Anaethalion knorri</i>	UJ	150	17.2	(59)
	† <i>Daitingichtys tischlingeri</i>	UJ	150	30.0	(10)
	† <i>Araripichthys castilhoi</i>	LK	146	42.0	(10)
	<i>Denticeps clupeoides</i>	R	0	15.0	(3)
	† <i>Clupea humilis</i>	EC	56	36.0	(3)
	† <i>Ellimma branneri</i>	LK	146	10.2	(63)
	† <i>Ellimmichthys longicostatus</i>	LK	146	10.4	(64)
	† <i>Ellimmichthys goodi</i>	LK	146	13.4	(64)
	† <i>Diplomystus dentatus</i>	UK	100	28.0	(10)
	† <i>Diplomystus shengliensis</i>	LK	146	5.5	(63)
	† <i>Paraclupea chetungensis</i>	LK	146	9.0	(64)
	† <i>Palaeodenticeps tanganyikae</i>	OC	34	3.0	(10)
	† <i>Engraulis eurystole</i>	EC	54	15.5	(59)
	† <i>Knighthia eocaena</i>	EC	57	9.0	(26)
	† <i>Santanacupea silvasantosi</i>	LK	125	12.0	(65)

Appendix 1. Continued

Class	Taxon	Horizon	Min. Age	TL (cm)	Reference
	† <i>Protoclupea chilensis</i>	UJ	150	9.7	(59)
	† <i>Aethalionopsis robusta</i>	UK	100	17.0	(10)
	† <i>Chanoïdes macropoma</i>	EC	50	12.0	(10)
	<i>Chanos chanos</i>	EC	54	170.0	(3)
	† <i>Chanos zignii</i>	UK	100	39.0	(10)
	† <i>Charitosomus hekelensis</i>	UK	100	14.0	(10)
	† <i>Notogoneus osculus</i>	EC	54	58.0	(10)
	† <i>Parachanos aethiopicus</i>	UK	100	22.0	(10)
	† <i>Tischlingerichthys vlohli</i>	UJ	150	12.8	(59)
	† <i>Santanichthys diasii</i>	UK	100	6.8	(65, 66)
	† <i>Amyzon aggregatus</i>	EC	54	22.0	(10)
	† <i>Esox lepidotus</i>	PC	61	70.0	(10)
	† <i>Umbrakramerii</i>	EC	54	17.0	(3)
	† <i>Umbraperpusilla</i>	EC	54	3.0	(10)
	<i>Dallia pectoralis</i>	R	0	33.0	(3)
	† <i>Estesesox tiemani</i>	PC	61	30.0	(67)
	† <i>Beurlenichthys ouricuriensis</i>	LK	125	6.0	(65, 66)
	† <i>Boltyshia brevicauda</i>	PC	61	8.0	(10)
	† <i>Prostomias maroccanus</i>	LK	125	17.0	(10)
	† <i>Dactyloporus grandis</i>	UK	95	44.0	(10)
	† <i>Hakelia laticauda</i>	UK	95	9.0	(10)
	† <i>Leptosoma elongatus</i>	UK	95	8.0	(10)
	† <i>Nematonotus longispinnus</i>	UK	95	14.0	(10)
	† <i>Tachynectes longipes</i>	UK	95	27.0	(10)
	† <i>Sardinoides monasterii</i>	UK	95	18.0	(10)
	† <i>Digoria ambigua</i>	UK	95	7.0	(10)
	† <i>Dinopteryx spinosus</i>	UK	95	14.0	(10)
	† <i>Aipichthys vilifer</i>	UK	95	8.0	(10)
	† <i>Pycnosteroides</i>	UK	95	7.0	(10)
	† <i>Stichocentrus livatus</i>	UK	95	6.0	(10)
	† <i>Enchodus macropterus</i>	UK	95	23.0	(10)
	† <i>Leptecodon rectus</i>	UK	95	23.0	(10)
	† <i>Palaeolycus dregensis</i>	UK	95	40.0	(10)
	† <i>Halec microlepis</i>	UK	95	18.0	(10)
	† <i>Phylactocephalus microlepis</i>	UK	95	15.0	(10)
	† <i>Apateopholis laniatus</i>	UK	95	31.0	(10)
	† <i>Ichthyotringa furcata</i>	UK	95	15.0	(10)
	† <i>Sardinus cordieri</i>	UK	95	14.0	(10)
	† <i>Ctenocephalichthys loreti</i>	UK	95	5.0	(10)
	† <i>Myripristis homopterygius</i>	ME	49	11.0	(10)
	† <i>Paraspinus cupulus</i>	UK	95	16.0	(10)
	† <i>Stichoberyx polydesmus</i>	LK	125	8.0	(10)
	† <i>Acrogaster heckeli</i>	UK	95	7.0	(10)
	† <i>Hoplopteryx antiquus</i>	UK	95	13.0	(10)
	† <i>Libanoberyx spinosus</i>	UK	95	5.5	(10)
	† <i>Lissoberyx arambourgi</i>	UK	95	6.0	(10)
	† <i>Stichopteryx lewisi</i>	UK	95	7.0	(10)
	† <i>Tubantia cataphractus</i>	UK	95	16.0	(10)
	† <i>Sphenocephalus fissicaudus</i>	UK	95	11.0	(10)
	† <i>Ctenothrissa vexillifer</i>	UK	95	9.0	(10)
	† <i>Mconichthys longipinnis</i>	UK	95	33.0	(10)
	† <i>Humilichthys orientalis</i>	UK	95	4.5	(10)
	† <i>Pattersonichthys delicatus</i>	UK	95	4.0	(10)
	† <i>Phonicolepis arcuatus</i>	UK	95	5.0	(10)
	† <i>Omosoma sahelalmae</i>	UK	95	8.0	(10)
	† <i>Platycornus germanus</i>	UK	95	20.0	(10)
	† <i>Pycnosterinx russeggeri</i>	UK	95	5.0	(10)

References

1. S. C. Morris, *The Crucible of Creation: The Burgess Shale and the Rise of Animals* (Oxford University Press, New York, 1998), pp.
2. B. W. Oelofsen, J. C. Loock, *South African Journal of Science* **77** (1981).
3. R. Froese, D. Pauly, *FishBase 2004: Concepts, Design and Data Sources* (ICLARM, Los Banos, 2004), pp.
4. D. Shu, X. Zhang, L. Chen, *Nature* **380** (1996).
5. J. Y. Chen, D. Y. Huang, C. W. Li, *Nature* **402**, 518 (Dec 2, 1999).
6. J. Mallatt, J. Y. Chen, *Journal of Morphology* **258**, 1 (2003).
7. D. G. Shu *et al.*, *Nature* **421**, 526 (2003).
8. P. Janvier, *Nature* **402**, 21 (Nov 4, 1999).
9. P. Janvier, *Early Vertebrates*, Oxford Monographs in Geology and Geophysics (Oxford University Press, Oxford, 1996), pp. 393.
10. K. A. Frickhinger, *Fossil Atlas Fishes* (Hans A Baensch, Melle, 1995), pp. 1088.
11. P. Janvier, R. Lund, *J. Vertebrate Paleontology* **2** (1983).
12. P. Y. Gagnier, in *Fosiles y Facies de Bolivia: v. 1 Vertebrados* S.-S. R., Ed. (Santa-Cruz, 1991), pp. 10.
13. G. C. Young, *J. Vert. Paleontol.* **25**, 261 (2005).
14. R. L. Carroll, *Vertebrate Paleontology and Evolution* (W. H. Freeman and company, New York, 1988), pp. 698.
15. J. A. Moy-Thomas, R. S. Miles, *Paleozoic Fishes* (Chapman and Hall, London, 1971), pp. 259.
16. J. M. Adrain, M. V. H. Wilson, *Journal of Vertebrate Paleontology* **14**, 301 (1994).
17. R. H. Denison, *Acanthodii. Handbook of Paleichthyology* (Gustav Fisher Verlag, Stuttgart, 1979), pp.
18. W. Van der Bruggen, P. L. Janvier, *Nature* **364**, 107 (1993).
19. K. L. Soehn, T. Märss, M. W. Caldwell, M. V. H. Wilson, *Journal of Vertebrate Paleontology* **21**, 651 (2001).
20. T. Marss, A. Ritchie, *Trans. Roy. Soc. Edinburgh, Earth Sci.* **88**, 143 (1998).
21. G. C. Young, G. R. Zhang, *J. Vert. Paleontol.* **16**, 623 (1996).
22. Z. Johanson, *Rec. Aus. Mus.* **50** (1998).
23. G. F. Hanke, M. V. H. Wilson, in *Recent Advances in the Origin and Early radiation of Vertebrates* G. Arratia, M. V. H. Wilson, R. Clouthier, Eds. (Verlag Dr. Friedrich Pfeil, Munchen, 2004), pp. 189–216.
24. M. J. Benton, *Vertebrate Palaeontology, 3rd.* (Blackwell, Oxford, 2005), pp.
25. R. Lund, W. L. Lund, *Geobios* **17**, 237 (1984).
26. J. G. Maisey, *Discovering Fossil Fishes* (Henry Holt & Co., New York, 1996), pp.
27. R. F. Miller, R. Clouthier, S. Turner, *Nature* **425**, 501 (2003).
28. J. A. Long, *The Rise of Fishes: 500 million years of evolution* (Johns Hopkins University Press, Baltimore & London, 1995), pp. 223.
29. H. Cappelletta, *Chondrichthyes II: Mesozoic and Cenozoic Elasmobranchii*, Handbook of Palaeoichthyology (Gustav Fischer Verlag, Stuttgart and New York: 1987), pp.
30. G. F. Hanke, M. V. H. Wilson, L. A. Lindo, *Can. J. Earth Sci.* **3811**, 1517 (2001).
31. G. F. Hanke, *Can. J. Earth Sci.* **39**, 1071 (2002).
32. J. A. Long, *J. Vert. Paleontol.* **21**, 815 (2001).
33. M. Zhu, X. B. Yu, *Nature* **418**, 767 (Aug 15, 2002).
34. R. Clouthier, in *Devonian Fishes and Plants of Miguasha, Quebec, Canada* H. P. Schultze, R. Clouthier, Eds. (Verlag Dr. Friedrich Pfeil, München, 1996), pp. 227–247.
35. R. Cloutier, P. E. Ahlberg, in *Interrelationships of Fishes* M. L. e. a. Stiassny, Ed. (–Academic Press, , San Diego, 1996), pp. 445–479.
36. R. Clouthier, P. L. Forey, *Environmental Biology of Fishes* **32**, 59 (1991).
37. R. Cloutier, *Environmental Biology of Fishes* **32**, 1 (1991).
38. R. H. Denison, *Fieldiana: Geology* **17**, 353 (1968).
39. R. Clouthier, *Geodiversitas* **19** (1997).
40. J. Saxon, *The Fossil Fishes of the North of Scotland* (Caithness Books, Thurso, 1975), pp.
41. A. Kemp, in *Mesozoic Fishes – Systematics and Paleoecology*, G. Arratia, G. Viohl, Eds. (Verlag Dr. Friedrich Pfeil, München, 1996), pp. 409–416.
42. C. S. Churcher, G. De iulii, *Palaeontology* **44**, 305 (2001).
43. G. Arratia, H. P. Schultze, J. Casciotta, *J. Morphology* **250**, 101 (2001).
44. M. Laurin, *Systematic Biology* **53**, 594 (Aug, 2004).
45. H. Blom, *Palaeontology* **48**, 111 (2005).
46. M. Zhu, X. B. Yu, P. Janvier, *Nature* **397**, 607 (Feb 18, 1999).
47. I. A. Hurley *et al.*, *Proc. R. Soc. B* **274**, 489 (2007).
48. G. Arratia, R. Clouthier, in *Recent Advances in the Origin and Early Radiation of Vertebrates* G. Arratia, M. V. H. Wilson, R. Cloutier, Eds. (Verlag Dr. Friedrich Pfeil, München, 2004), pp. 583–598.
49. G. Arratia, R. Cloutier, *Journal of Vertebrate Paleontology* **22**, 33A (2002).
50. M. Friedman, H. Blom, *J. Paleontology* **80**, 1186 (2006).
51. R. Lund, *Geodiversitas* **22**, 171 (2000).
52. Anonymous, <http://www.fossilweb.de/perm/> (2007).
53. S. Štamberg, *Journal of the Czech Geological Society* **47**, 147 (2002).
54. M. Gayet, F. J. Meunier, C. Werner, *Palaeontology* **45**, 361 (2002).
55. M. I. Coates, *Zoological Journal of Linnean Society* **122**, 27 (1998).
56. W. E. Bemis, E. K. Findeis, L. Grande, *Env. Biol. Fishes* **48**, 25 (1997).
57. A. Tintori, in *Systematics and Paleoecology* G. Arratia, G. Viohl, Eds. (Verlag Dr. Friedrich Pfeil, München, 1996), pp. 167–179.

58. L. Grand, W. E. Bemis, *Journal of Vertebrate Paleontology* **18**, 1 (1998).
 59. G. Arratia, *Palaeo Ichthyologica* **7**, 1 (1997).
 60. G. Arratia, R. A. Scasso, W. Kiessling, *Journal of Vertebrate Paleontology* **24**, 1 (2004).
 61. J. G. Lundberg, C. B., *Biotropica* **24**, 2 (1992).
 62. A. Murray, M. Wilson, *Zoological Journal of the Linnean Society* **144**, 213 (2005).
 63. M. M. Chang, J. G. Maisey, *American Museum Novitates* **3404**, 1 (2003).
 64. M. M. Chang, L. Grande, *Fieldiana Geology New Series*, **37**, 1 (1997).
 65. J. G. Maisey, *Santana Fossils: An Illustrated Atlas*, Cretaceous Research (Neptune City, 1991), pp. 459.
 66. A. Filleul, J. G. Maisey, *American Museum Novitates* **3455**, 1 (2004).
 67. M. V. H. Wilson, D. B. Brinkman, A. G. Neuman, *Journal of Paleontology* **66**, 839 (1992).
 68. F. J. de Figueiredo, V. Gallo, *Boletim do Museu Nacional, Geologia* **73**, 1 (2004).

Appendix 2. The extant fish dataset. Size (average standard length in cm and ln cm), size variation (standard deviation, skew, kurtosis) and species richness (N) for all 515 recognized extant fish families. Data summaries of maximum recorded standard lengths for 24,259 species from (Froese and Pauly 2005). NA, not applicable. Taxa sorted alphabetically by species.

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Abyssocottidae	13	15.0	2.6	0.4	-0.2	0.7
Acanthuridae	79	38.1	3.5	0.5	0.0	-0.6
Acestrorhynchidae	15	19.1	2.8	0.6	-0.3	-1.6
Achiridae	31	14.5	2.6	0.5	-0.8	-0.1
Achiropsettidae	6	33.1	3.2	0.9	-1.0	-0.9
Acipenseridae	25	239.3	5.3	0.7	-0.6	1.0
Adrianiichthyidae	25	5.1	1.4	0.6	0.8	0.8
Agonidae	37	18.3	2.8	0.4	0.0	-0.4
Akysidae	35	6.9	1.8	0.5	0.4	-0.7
Albulidae	6	75.8	4.3	0.4	-0.7	-1.3
Alepisauridae	2	155.5	5.0	0.6	-	-
Alepocephalidae	78	32.7	3.4	0.4	0.3	0.3
Alestidae	107	16.2	2.4	0.8	0.9	0.9
Alopiidae	3	543.7	6.3	0.3	0.8	-
Amarsipidae	1	12.0	2.5	-	-	-
Ambassidae	45	9.4	2.1	0.5	-0.8	0.9
Amblycipitidae	14	8.5	2.1	0.4	0.2	-1.5
Amblyopsidae	6	8.2	2.1	0.2	0.3	-1.2
Amiidae	1	109.0	4.7	-	-	-
Ammodytidae	23	18.5	2.8	0.4	0.2	-0.3
Amphiliidae	65	8.9	2.0	0.6	0.0	0.4
Anabantidae	35	11.7	2.3	0.6	0.2	-0.8
Anablepidae	11	14.2	2.3	0.8	0.4	-1.2
Anacanthobatidae	5	34.0	3.5	0.3	-0.7	-2.3
Anarhichadidae	5	170.0	5.1	0.3	-0.8	1.3
Anguillidae	18	137.6	4.9	0.3	-0.4	0.3
Anomalopidae	6	13.9	2.4	0.7	0.8	2.0
Anoplogasteridae	2	10.6	2.3	0.7	-	-
Anoplopomatidae	2	151.5	5.0	0.3	-	-
Anostomidae	72	21.4	2.9	0.5	-0.1	-0.4
Anotopteridae	3	115.7	4.7	0.2	1.4	-
Antennariidae	44	15.0	2.5	0.6	0.1	-0.7
Aphredoderidae	1	14.0	2.6	-	-	-
Aphyonidae	11	9.4	2.2	0.4	0.4	0.4
Apistidae	2	19.5	3.0	0.0	-	-
Aploactinidae	2	6.0	1.5	1.1	-	-
Aplocheilidae	225	5.8	1.7	0.2	0.3	0.7
Aplodactylidae	5	41.1	3.6	0.5	-1.3	2.2
Apogonidae	281	8.8	2.1	0.5	-0.1	-0.4
Apteronotidae	46	33.5	3.4	0.4	1.3	2.9
Arapaimidae	2	275.0	5.4	1.1	-	-
Argentinidae	20	19.2	2.8	0.5	1.0	2.5

Appendix 2. Continued

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Ariidae	122	48.1	3.7	0.5	0.5	0.5
Ariommatidae	8	38.0	3.5	0.5	0.9	0.1
Arripidae	4	77.8	4.3	0.4	-1.9	3.7
Artedidraconidae	27	19.2	2.9	0.5	-1.3	1.8
Aspredinidae	36	9.5	1.9	0.8	0.0	-0.4
Astroblepidae	53	8.8	2.1	0.4	0.1	2.9
Ateleopodidae	6	127.0	4.7	0.7	-0.6	-1.3
Atherinidae	54	9.7	2.2	0.4	0.1	0.4
Atherinopsidae	41	15.9	2.6	0.6	0.6	0.1
Auchenipteridae	76	14.2	2.4	0.7	-0.2	-0.2
Aulopidae	8	32.3	3.4	0.4	0.8	-0.8
Aulorhynchidae	2	15.4	2.7	0.2	-	-
Aulostomidae	3	85.0	4.4	0.2	1.4	-
Badidae	16	4.0	1.3	0.4	-0.1	0.4
Bagridae	219	30.3	3.0	0.8	0.2	0.0
Balistidae	39	41.4	3.6	0.5	-0.4	0.0
Balitoridae	416	7.5	1.9	0.5	0.6	1.8
Barbourisiidae	1	34.5	3.5	-	-	-
Bathyclupeidae	2	19.5	3.0	0.1	-	-
Bathydraconidae	15	26.9	3.2	0.4	0.6	0.2
Bathylagidae	19	16.4	2.8	0.3	0.1	-0.5
Bathylutichthyidae	1	10.0	2.3	-	-	-
Bathymasteridae	6	23.3	3.1	0.4	0.6	-1.0
Bathysauroididae	1	29.0	3.4	-	-	-
Batrachoididae	62	25.1	3.1	0.6	-0.8	0.3
Bedotiidae	11	8.1	2.0	0.4	-0.2	-0.6
Belonidae	43	68.7	4.0	0.8	-1.0	1.7
Bembridae	6	22.7	3.1	0.4	-1.7	3.1
Berycidae	9	50.8	3.8	0.5	-0.3	-0.9
Blenniidae	325	8.7	2.0	0.5	0.2	0.0
Bothidae	117	16.2	2.7	0.5	-0.2	0.3
Bovichtidae	4	33.6	3.2	0.9	0.7	0.4
Brachaeluridae	2	99.0	4.6	0.3	-	-
Brachionichthyidae	2	11.5	2.4	0.4	-	-
Bramidae	19	51.4	3.8	0.6	-1.5	3.6
Bregmacerotidae	12	6.4	1.7	0.7	-1.2	0.7
Bythitidae	61	16.5	2.5	0.8	0.8	0.1
Caesionidae	20	29.4	3.3	0.4	-0.1	-0.3
Callanthiidae	9	26.1	3.1	0.7	0.2	-0.7
Callichthyidae	173	5.2	1.6	0.4	0.9	3.4
Callionymidae	123	10.7	2.1	0.7	-0.5	-0.7
Callorhynchidae	3	112.1	4.7	0.2	-1.7	-
Caproidae	8	15.7	2.6	0.6	0.0	-0.9
Caracanthidae	4	4.5	1.5	0.3	-2.0	4.0
Carangidae	141	68.9	4.1	0.5	0.2	-0.3
Carapidae	33	19.8	2.9	0.4	-0.1	-0.7
Carcharhinidae	49	220.3	5.2	0.6	0.0	-0.6
Caristiidae	4	27.6	3.3	0.3	-1.9	3.7
Catostomidae	61	53.8	3.9	0.5	-0.5	-0.4
Caulophryniidae	3	16.4	2.8	0.2	1.6	-
Centracanthidae	8	28.4	3.3	0.2	-0.4	-1.0
Centrarchidae	30	37.1	3.4	0.7	-0.5	-0.5
Centriscidae	12	22.0	3.0	0.3	0.4	-1.7
Centrogeniidae	1	25.0	3.2	-	-	-
Centrolophidae	25	63.7	4.0	0.7	-0.6	-0.5
Centrophoridae	14	117.3	4.7	0.3	0.2	-1.3
Centrophryniidae	1	23.0	3.1	-	-	-
Centropomidae	22	85.9	4.3	0.6	0.2	-1.2
Cepolidae	15	41.3	3.6	0.5	-1.0	2.0

Appendix 2. Continued

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Ceratiidae	4	69.0	4.1	0.7	-0.4	-2.1
Ceratodontidae	1	170.0	5.1	–	–	–
Cetomimidae	13	13.9	2.5	0.6	0.2	-1.0
Cetopsidae	23	10.8	2.1	0.8	-0.2	-0.8
Cetorhinidae	1	900.0	6.8	–	–	–
Chacidae	3	20.0	3.0	0.0	–	–
Chaenopsidae	69	6.3	1.7	0.5	0.8	1.6
Chaetodontidae	125	17.1	2.8	0.2	0.0	0.2
Champsodontidae	12	11.2	2.4	0.2	-0.2	-0.5
Chanidae	1	124.0	4.8	–	–	–
Channichthyidae	19	45.4	3.8	0.3	-0.1	-1.2
Channidae	28	48.6	3.6	0.7	0.6	-0.7
Characidae	842	9.7	1.9	0.8	0.7	0.3
Chaudhuriidae	9	5.5	1.7	0.3	-0.8	-0.3
Chaunacidae	14	21.6	3.0	0.4	-0.1	-1.2
Cheilodactylidae	23	56.2	3.9	0.6	0.5	0.2
Cheimarrichthyidae	1	15.0	2.7	–	–	–
Chiasmodontidae	13	18.7	2.8	0.7	-1.8	2.2
Chilodontidae	7	10.2	2.3	0.4	0.6	-1.5
Chimaeridae	17	93.3	4.5	0.4	-0.3	-0.8
Chirocentridae	2	100.0	4.6	0.0	–	–
Chironemidae	3	31.7	3.4	0.4	-1.5	–
Chlamydoselachidae	1	200.0	5.3	–	–	–
Chlopsidae	21	21.1	3.0	0.3	0.3	-0.4
Chlorophthalmidae	9	24.3	3.1	0.3	0.0	0.0
Cichlidae	1456	15.2	2.6	0.6	0.2	0.3
Cirrhitidae	28	15.5	2.6	0.5	1.0	1.2
Citharidae	6	25.6	3.2	0.3	-0.9	0.1
Citharinidae	102	19.9	2.3	1.1	0.4	-1.1
Clariidae	102	40.8	3.4	0.8	0.1	-0.2
Clinidae	79	14.5	2.5	0.6	0.5	0.3
Clupeidae	202	20.0	2.7	0.7	-0.2	-0.3
Cobitidae	137	11.5	2.2	0.6	0.5	0.1
Coiidae	5	37.4	3.6	0.2	-0.3	-2.1
Colocongridae	5	56.9	4.0	0.3	1.0	2.2
Comephoridae	2	18.5	2.9	0.2	–	–
Congiopodidae	6	39.9	3.6	0.5	-0.2	0.9
Congridae	121	57.9	3.9	0.6	-0.1	2.7
Coryphaenidae	2	168.5	5.1	0.4	–	–
Cottidae	190	16.3	2.6	0.7	0.3	0.2
Cottocomephoridae	6	19.8	3.0	0.2	-1.1	1.8
Cranoglanididae	2	36.0	3.6	0.3	–	–
Creediidae	16	5.1	1.6	0.4	-0.3	-0.6
Crenuchidae	75	4.7	1.5	0.5	-0.3	-0.2
Cryptacanthodidae	3	79.3	4.2	0.7	-1.5	–
Ctenoluciidae	7	7.0	3.6	3.6	0.4	1.5
Curimatidae	98	12.6	2.4	0.5	-0.6	-0.1
Cyclopteridae	19	13.3	2.2	0.8	0.8	1.4
Cyematidae	2	15.5	2.7	0.0	–	–
Cynodontidae	14	36.2	3.3	0.7	0.4	-0.6
Cynoglossidae	113	21.0	2.9	0.6	0.1	-0.1
Cyprinidae	1895	21.2	2.6	0.9	0.4	-0.1
Cyprinodontidae	120	7.0	1.8	0.4	1.2	1.5
Cyttidae	3	45.0	3.8	0.2	1.7	–
Dactylopteridae	7	35.2	3.4	0.7	0.1	1.8
Dactyloscopidae	36	5.9	1.7	0.4	0.1	0.4
Dalatiidae	64	78.2	4.0	0.7	1.2	2.3
Dasyatidae	60	136.4	4.6	0.8	-0.3	-0.4
Dentatherinidae	1	5.0	1.6	–	–	–

Appendix 2. Continued

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Denticipitidae	1	15.0	2.7	–	–	–
Derichthyidae	3	43.3	3.7	0.3	0.5	–
Diceratiidae	5	17.2	2.8	0.4	0.0	–2.8
Dichistiidae	2	57.5	4.0	0.6	–	–
Dinolestidae	1	84.0	4.4	–	–	–
Dinopercidae	2	52.9	3.9	0.6	–	–
Diodontidae	20	41.4	3.7	0.4	0.7	–0.4
Diplomystidae	6	24.2	3.2	0.2	–0.3	1.2
Diretmidae	4	31.6	3.4	0.3	–0.1	–4.6
Doradidae	73	22.8	2.8	0.7	0.7	0.1
Draconettidae	10	9.8	2.3	0.2	–1.4	3.5
Drepaneidae	3	48.3	3.9	0.1	–1.7	–
Echeneidae	8	66.7	4.1	0.5	–0.5	–1.3
Echinorhinidae	2	218.0	5.3	0.6	–	–
Elassomatidae	5	3.6	1.3	0.2	2.1	4.5
Eleginopidae	1	60.0	4.1	–	–	–
Eleotridae	131	15.4	2.5	0.7	0.1	–0.3
Elopidae	6	99.8	4.6	0.1	1.1	1.8
Embiotocidae	22	28.1	3.3	0.4	–0.1	–1.1
Emmelichthyidae	16	35.0	3.4	0.6	–0.4	0.1
Engraulidae	139	13.8	2.5	0.6	–0.3	0.2
Enoplosidae	1	50.0	3.9	–	–	–
Ephippidae	15	46.3	3.7	0.5	–0.3	0.1
Epigonidae	20	19.4	2.7	0.7	–0.2	1.5
Erethistidae	10	7.2	1.6	0.8	1.5	2.3
Ereuniidae	3	25.8	3.2	0.2	–1.4	–
Erythrinidae	11	46.5	3.6	0.7	0.3	–1.3
Eschmeyeridae	1	4.1	1.4	–	–	–
Esocidae	6	101.5	4.5	0.7	–0.4	–1.8
Euclichthyidae	1	35.0	3.6	–	–	–
Eurypharyngidae	1	100.0	4.6	–	–	–
Evermannellidae	5	15.0	2.7	0.2	0.1	–4.6
Exocoetidae	57	26.7	3.2	0.3	–0.1	–0.3
Fistulariidae	4	157.5	5.0	0.5	–1.8	3.0
Fundulidae	45	9.4	2.2	0.4	0.1	0.1
Gadidae	24	65.8	4.0	0.7	0.0	–0.2
Galaxiidae	45	13.0	2.4	0.6	0.5	–0.4
Gasteropelecidae	9	4.4	1.4	0.5	0.4	–1.3
Gasterosteidae	10	9.3	2.2	0.4	1.7	3.9
Gempylidae	24	78.9	4.1	0.8	0.4	–1.3
Geotriidae	4	35.8	3.4	0.7	0.1	–5.6
Gerreidae	50	21.9	3.0	0.4	–0.4	0.3
Gibberichthyidae	2	12.5	2.5	0.1	–	–
Gigantactinidae	19	18.2	2.7	0.7	–0.5	0.1
Giganturidae	2	18.0	2.9	0.2	–	–
Ginglymostomatidae	3	275.0	5.4	0.9	–1.5	–
Glaucosomatidae	4	67.3	4.1	0.6	0.4	–0.9
Gnathanacanthidae	1	30.0	3.4	–	–	–
Gobiesocidae	114	5.4	1.5	0.6	0.3	0.2
Gobiidae	1070	7.6	1.8	0.7	0.3	0.0
Gonorynchidae	5	43.9	3.7	0.3	–0.3	–2.7
Gonostomatidae	28	12.5	2.2	0.8	0.3	–1.0
Goodeidae	48	6.8	1.9	0.3	0.4	0.1
Grammatidae	12	4.7	1.4	0.6	–0.2	–0.8
Grammicolepididae	2	39.5	3.4	1.0	–	–
Gymnarchidae	1	167.0	5.1	–	–	–
Gymnotidae	25	34.0	3.2	0.7	1.4	4.1
Gymnuridae	9	167.6	4.9	0.8	–0.5	0.4
Gyrinocheilidae	3	30.5	3.4	0.1	1.7	–

Appendix 2. *Continued*

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Haemulidae	135	45.1	3.7	0.5	-0.2	-0.1
Halosauridae	13	54.5	4.0	0.2	0.4	1.3
Harpagiferidae	6	8.2	2.1	0.1	0.0	-1.9
Hemigaleidae	7	117.5	4.7	0.5	0.1	1.6
Hemiodontidae	28	16.7	2.7	0.4	-0.1	-1.0
Hemiramphidae	101	19.8	2.8	0.7	-0.3	-0.7
Hemiscyllidae	13	74.5	4.3	0.2	0.1	-0.1
Hemitripterae	8	31.5	3.2	0.9	-0.1	-1.0
Hepsetidae	1	70.0	4.2	–	–	–
Heptapteridae	70	12.1	2.3	0.6	0.1	-0.1
Heterenchelyidae	7	69.3	4.1	0.5	0.7	-0.1
Heterodontidae	8	118.0	4.7	0.3	-0.6	-0.3
Heteropneustidae	3	24.0	3.1	0.4	-1.6	–
Hexagrammidae	12	52.4	3.8	0.5	0.9	1.2
Hexanchidae	4	275.5	5.5	0.5	0.4	-1.9
Hexatrygonidae	4	103.8	4.6	0.4	-1.2	2.2
Himantolophidae	7	22.6	2.7	1.0	0.0	-1.4
Hiodontidae	2	49.5	3.9	0.1	–	–
Hispidoberycidae	1	18.1	2.9	–	–	–
Holocentridae	81	23.2	3.1	0.4	-0.1	0.6
Hoplichthyidae	8	21.9	3.0	0.5	-0.8	2.3
Hypopomidae	14	18.7	2.8	0.4	0.0	-0.6
Hypoptychidae	1	6.7	1.9	–	–	–
Icosteidae	1	213.0	5.4	–	–	–
Ictaluridae	40	31.0	2.9	0.9	0.9	0.0
Indostomidae	3	2.8	1.0	0.1	-1.7	–
Inermiidae	2	18.0	2.9	0.4	–	–
Ipnopidae	20	22.5	3.1	0.4	0.1	-1.1
Istiophoridae	11	341.5	5.8	0.3	-0.1	-1.2
Kneriidae	30	6.6	1.8	0.4	-0.8	1.7
Kraemeriidae	4	4.1	1.4	0.1	-1.0	-0.7
Kuhliidae	12	22.9	3.0	0.4	0.5	0.0
Kurtidae	2	37.8	3.3	1.1	–	–
Kyphosidae	41	43.5	3.7	0.5	-0.2	-1.0
Labridae	536	26.8	3.0	0.7	0.4	-0.4
Labrisomidae	79	7.1	1.8	0.6	0.1	-0.6
Lactariidae	1	40.0	3.7	–	–	–
Lamnidae	5	438.4	6.0	0.3	1.5	2.7
Lampridae	2	155.0	5.0	0.4	–	–
Lateolabracidae	2	98.0	4.6	0.1	–	–
Latimeriidae	2	154.0	5.0	0.1	–	–
Latridae	4	76.3	4.3	0.5	-0.2	0.5
Lebiasinidae	60	7.3	1.8	0.6	0.2	-1.0
Leiognathidae	34	12.8	2.5	0.4	0.1	-0.1
Lepidogalaxiidae	1	6.7	1.9	–	–	–
Lepidosirenidae	1	125.0	4.8	–	–	–
Lepisosteidae	6	142.8	4.9	0.3	0.1	-1.2
Leptobramidae	1	37.5	3.6	–	–	–
Leptochariidae	1	82.0	4.4	–	–	–
Leptochilichthyidae	3	28.5	3.3	0.1	-1.7	–
Leptoscopidae	3	13.3	2.6	0.2	1.5	–
Lethrinidae	37	53.4	3.9	0.4	-0.4	-0.1
Linophrynidae	18	7.6	1.9	0.5	1.0	0.8
Liparidae	218	15.6	2.5	0.7	0.0	-0.1
Lobotidae	2	105.0	4.7	0.1	–	–
Lophichthyidae	1	5.1	1.6	–	–	–
Lophiidae	23	52.9	3.6	0.8	0.3	-0.2
Lophotidae	3	183.3	5.2	0.2	-1.7	–
Loricariidae	670	16.2	2.6	0.7	-0.1	-0.1
Lotidae	20	62.0	3.8	0.7	0.8	-0.6

Appendix 2. Continued

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Lutjanidae	108	66.2	4.1	0.5	-0.2	-0.3
Macrouridae	300	39.8	3.6	0.4	-0.1	0.3
Malacanthidae	41	39.6	3.5	0.6	0.0	-0.7
Malapteruridae	11	47.4	3.6	0.8	0.2	-1.3
Mastacembelidae	71	30.3	3.3	0.5	0.2	0.5
Megachasmidae	1	549.0	6.3	–	–	–
Megalomycteridae	3	4.1	1.4	0.1	0.1	–
Megalopidae	2	200.0	5.3	0.4	–	–
Melamphidae	25	7.9	1.9	0.6	-0.7	-0.4
Melanocetidae	4	13.2	2.6	0.2	1.8	3.5
Melanonidae	2	23.4	3.1	0.3	–	–
Melanotaeniidae	67	8.7	2.1	0.3	-0.1	0.4
Menidae	1	30.0	3.4	–	–	–
Merlucciidae	20	87.3	4.4	0.5	-1.0	0.1
Microdesmidae	40	9.1	2.1	0.5	-0.3	0.0
Microstomatidae	19	16.8	2.8	0.4	-0.2	-1.4
Mirapinnidae	4	4.7	1.5	0.2	-1.3	1.0
Mochokidae	188	19.3	2.6	0.8	-0.1	-0.9
Molidae	5	256.0	5.5	0.5	-1.6	2.1
Monacanthidae	100	24.0	2.9	0.7	-0.1	-0.1
Monocentridae	3	16.3	2.7	0.4	-1.0	–
Monodactylidae	6	22.1	3.0	0.5	-2.0	4.4
Monognathidae	15	7.1	1.9	0.3	0.7	-0.5
Moridae	86	29.9	3.2	0.6	-0.5	2.1
Moringuidae	8	64.0	4.0	0.6	0.7	-1.0
Mormyridae	200	23.7	2.9	0.7	0.2	-0.4
Moronidae	6	85.6	4.3	0.6	1.2	0.6
Mugilidae	72	44.3	3.6	0.6	-0.8	1.0
Mullidae	65	29.9	3.3	0.4	-0.8	0.7
Muraenesocidae	13	135.3	4.8	0.5	0.2	-1.9
Muraenidae	171	79.2	4.1	0.7	-0.3	0.4
Muraenolepididae	4	33.8	3.5	0.1	0.7	-1.9
Myctophidae	203	9.3	2.1	0.5	0.0	-0.3
Myliobatidae	33	183.2	5.0	0.6	0.7	0.8
Myrocongridae	4	44.1	3.8	0.2	0.3	-3.7
Myxinidae	69	51.2	3.9	0.4	-0.3	0.1
Nandidae	8	13.1	2.5	0.5	-0.6	-1.2
Narcinidae	27	36.2	3.5	0.4	-0.3	0.2
Nematistiidae	1	163.0	5.1	–	–	–
Nemichthyidae	7	102.9	4.6	0.4	0.1	-1.7
Nemipteridae	63	22.5	3.1	0.3	-0.7	0.7
Neoceratiidae	1	6.0	1.8	–	–	–
Neoscopelidae	3	25.2	3.2	0.2	-0.2	–
Neosebastidae	8	29.9	3.2	0.7	-1.1	-0.1
Nettastomatidae	29	57.4	4.0	0.4	-0.3	0.1
Nomeidae	17	42.4	3.5	0.8	0.6	-1.2
Normanichthyidae	1	11.0	2.4	–	–	–
Notacanthidae	8	49.4	3.7	0.8	-0.6	0.3
Notocheiridae	5	5.9	1.7	0.3	0.2	-2.2
Notograptidae	2	10.3	2.3	0.0	–	–
Notopteridae	8	85.4	4.3	0.7	-0.9	-0.5
Notosudidae	14	25.8	3.2	0.4	-0.2	-0.1
Nototheniidae	42	41.8	3.5	0.7	0.9	1.1
Odacidae	12	22.7	3.0	0.5	0.0	-1.8
Odontaspidae	4	354.3	5.9	0.1	-1.8	3.4
Odontobutidae	10	10.8	2.2	0.5	0.4	-0.7
Ogcocephalidae	62	13.9	2.5	0.6	0.3	-0.4
Olyridae	4	10.9	2.4	0.3	-0.8	1.9
Omosudidae	1	23.0	3.1	–	–	–
Oneirodidae	46	10.6	2.2	0.6	-0.7	-0.3

Appendix 2. *Continued*

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Ophichthidae	207	61.6	3.9	0.6	-0.3	0.5
Ophidiidae	156	36.6	3.3	0.7	0.6	0.3
Opisthoproctidae	10	18.5	2.7	0.7	-0.2	1.1
Opistognathidae	43	20.2	2.6	0.9	0.3	-0.5
Oplegnathidae	7	71.0	4.2	0.2	-0.8	0.0
Orectolobidae	6	165.0	4.9	0.7	0.5	-1.6
Oreosomatidae	10	34.1	3.4	0.4	-0.2	-0.9
Osmeridae	15	27.7	3.2	0.5	0.6	0.6
Osphronemidae	88	9.2	1.9	0.7	1.1	1.8
Osteoglossidae	5	98.0	4.6	0.1	1.6	2.2
Ostraciidae	34	28.2	3.2	0.5	-0.1	-1.3
Ostracoberycidae	3	15.6	2.7	0.5	-0.8	-
Pangasiidae	27	92.8	4.2	0.8	0.3	-0.6
Pantodontidae	1	11.9	2.5	-	-	-
Parabembridae	2	19.6	2.9	0.3	-	-
Parabrotulidae	3	5.0	1.6	0.1	-0.1	-
Parakysidae	5	4.2	1.4	0.4	-1.0	1.0
Paralepididae	39	25.5	3.0	0.8	-0.6	1.3
Paralichthyidae	83	32.8	3.3	0.6	0.4	0.2
Parascorpididae	1	60.0	4.1	-	-	-
Parascylliidae	7	65.4	4.1	0.4	-0.5	-2.0
Paraulopidae	8	13.3	2.6	0.3	0.2	1.1
Parazenidae	3	17.6	2.7	0.7	0.5	-
Parodontidae	26	10.2	2.3	0.3	-0.9	0.3
Pataecidae	3	18.0	2.7	0.8	-1.5	-
Pegasidae	5	10.5	2.3	0.3	1.7	3.1
Pempheridae	23	16.1	2.7	0.3	-0.6	1.8
Pentacerotidae	12	53.5	3.9	0.5	0.3	-1.1
Percichthyidae	30	31.9	3.0	1.0	0.6	-0.9
Percidae	173	11.5	2.2	0.6	2.2	6.4
Perciliidae	2	9.3	2.2	0.0	-	-
Percophidae	33	17.0	2.6	0.7	-0.5	-0.5
Percopsidae	2	14.8	2.6	0.5	-	-
Peristediidae	31	26.3	3.1	0.5	-0.1	0.3
Petromyzontidae	36	29.9	3.2	0.5	1.2	1.4
Phallostethidae	19	2.4	0.9	0.3	0.3	0.1
Pholidae	14	24.4	3.1	0.4	-1.0	1.4
Pholidichthyidae	2	29.3	3.4	0.2	-	-
Photichthyidae	21	13.1	2.4	0.7	0.1	-1.2
Phractolaemidae	1	19.0	2.9	-	-	-
Phycidae	11	63.5	4.1	0.4	0.9	-0.1
Pimelodidae	56	60.5	3.7	0.9	0.0	-0.1
Pinguipedidae	50	24.4	3.0	0.6	1.3	2.2
Platycephalidae	61	35.0	3.4	0.6	0.0	0.0
Platytrictidae	37	19.3	2.9	0.3	-0.3	-0.6
Plecoglossidae	2	40.7	3.3	1.3	-	-
Plectrogenidae	2	9.5	2.2	0.4	-	-
Plesiobatidae	1	270.0	5.6	-	-	-
Plesiopidae	40	10.8	2.2	0.7	-0.2	-0.5
Pleuronectidae	89	46.8	3.6	0.7	0.1	0.2
Plotosidae	32	42.0	3.5	0.6	0.3	0.6
Poeciliidae	285	5.2	1.6	0.4	0.2	1.0
Polycentridae	2	9.0	2.2	0.2	-	-
Polymixiidae	10	26.7	3.2	0.4	0.8	-0.1
Polynemidae	40	41.7	3.3	0.8	1.1	0.8
Polyodontidae	2	260.5	5.6	0.2	-	-
Polyprionidae	5	202.0	5.3	0.2	-0.8	2.1
Polypteridae	18	52.4	3.9	0.4	0.0	-1.1
Pomacanthidae	84	21.9	2.9	0.6	0.0	-1.0
Pomacentridae	349	11.4	2.4	0.4	0.1	0.0

Appendix 2. *Continued*

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Pomatomidae	1	130.0	4.9	–	–	–
Potamotrygonidae	18	50.9	3.8	0.4	0.0	–0.2
Priacanthidae	18	34.3	3.5	0.3	0.6	0.4
Pristidae	7	558.0	6.2	0.6	–2.2	5.0
Pristigasteridae	22	25.6	3.0	0.7	–0.3	1.7
Pristiophoridae	5	129.0	4.8	0.3	–1.2	2.3
Prochilodontidae	21	32.8	3.5	0.2	0.3	–1.1
Profundulidae	5	9.0	2.2	0.2	0.8	–1.4
Proscylliidae	6	78.9	4.1	0.8	0.6	–0.4
Protopteridae	6	93.3	4.5	0.4	–1.7	4.0
Psettodidae	3	66.3	4.2	0.2	0.6	–
Pseudaphritidae	1	36.0	3.6	–	–	–
Pseudocarchariidae	1	110.0	4.7	–	–	–
Pseudochromidae	104	8.4	2.0	0.5	0.3	0.4
Pseudomugilidae	15	3.6	1.3	0.2	0.5	–1.1
Pseudopimelodidae	26	19.4	2.5	0.9	0.2	–0.7
Pseudotriakidae	1	295.0	5.7	–	–	–
Pseudotriconotidae	2	7.5	2.0	0.3	–	–
Psilorhynchidae	6	7.2	1.9	0.2	0.4	–1.6
Psychrolutidae	30	24.3	3.0	0.6	–0.7	0.1
Ptilichthyidae	1	34.0	3.5	–	–	–
Rachycentridae	1	200.0	5.3	–	–	–
Radiicephalidae	1	76.0	4.3	–	–	–
Rajidae	154	78.0	4.2	0.6	0.1	–0.2
Regalecidae	3	468.5	4.8	2.8	–1.3	–
Retropinnidae	6	14.7	2.5	0.6	1.0	–0.9
Rhamphichthyidae	7	50.8	3.7	0.7	–0.2	–1.7
Rhamphocottidae	1	8.9	2.2	–	–	–
Rhincodontidae	1	2000.0	7.6	–	–	–
Rhinobatidae	43	121.9	4.7	0.5	0.6	0.0
Rhinochimaeridae	7	104.6	4.6	0.3	–0.4	–2.5
Rhyacichthyidae	2	21.5	3.1	0.2	–	–
Rivulidae	225	6.1	1.7	0.4	0.3	0.2
Rondeletiidae	2	11.1	2.4	0.0	–	–
Saccopharyngidae	4	104.9	4.6	0.4	0.1	0.5
Salangidae	13	10.7	2.3	0.4	0.1	–1.7
Salmonidae	104	65.8	4.0	0.6	0.0	–0.4
Samaridae	17	10.8	2.3	0.4	0.0	–0.4
Scatophagidae	4	27.3	3.2	0.7	–1.9	3.6
Schilbeidae	56	28.9	3.1	0.7	0.4	–0.1
Schindleriidae	2	1.7	0.4	0.8	–	–
Sciaenidae	257	47.8	3.6	0.7	0.4	0.0
Scoloplacidae	4	1.6	0.4	0.2	–0.1	–3.4
Scomberesocidae	5	31.6	3.1	1.0	–1.0	–0.7
Scombridae	53	126.1	4.6	0.6	–0.2	0.1
Scombrobracidae	1	30.0	3.4	–	–	–
Scombropidae	3	124.7	4.8	0.2	0.6	–
Scopelarchidae	15	13.6	2.5	0.6	0.1	0.6
Scophthalmidae	9	46.9	3.7	0.7	–0.5	–0.3
Scorpaenidae	172	18.7	2.7	0.7	0.0	–0.7
Scyliorhinidae	108	63.2	4.0	0.5	0.8	2.8
Scytalinidae	1	15.0	2.7	–	–	–
Sebastidae	116	44.7	3.7	0.5	–0.2	–0.2
Serranidae	511	38.7	3.2	1.0	0.1	–0.6
Serrivomeridae	8	59.2	4.1	0.3	–0.6	–1.0
Setarchidae	4	18.5	2.9	0.3	0.0	1.5
Siganidae	27	32.5	3.4	0.3	0.2	–1.0
Sillaginidae	30	29.0	3.3	0.4	0.3	–0.3
Siluridae	84	45.8	3.2	1.0	0.8	0.8
Sisoridae	95	18.2	2.5	0.7	1.8	4.5

Appendix 2. Continued

Family	N	Avg. (cm)	Avg. (ln cm)	Stdev	Skew	Kurt
Soleidae	105	21.9	2.9	0.7	-0.2	0.9
Solenostomidae	4	11.2	2.3	0.5	-0.7	1.0
Sparidae	120	56.2	3.9	0.5	0.3	0.4
Sphyracidae	25	98.8	4.4	0.6	-0.2	-1.1
Sphyrnidae	9	285.1	5.5	0.7	0.2	-1.4
Squalidae	10	97.5	4.5	0.3	0.3	-0.3
Squatinae	13	136.5	4.8	0.4	-1.4	1.4
Stegostomatidae	1	235.0	5.5	–	–	–
Stephanoberycidae	2	10.6	2.3	0.3	–	–
Sternoptychidae	62	5.9	1.7	0.4	-0.2	0.5
Sternopygidae	28	40.8	3.6	0.5	0.2	1.3
Stichaeidae	68	21.8	2.9	0.6	0.3	-0.5
Stomiidae	225	18.6	2.8	0.5	-0.3	1.4
Stromateidae	17	32.6	3.4	0.4	0.2	0.5
Stylephoridae	1	28.0	3.3	–	–	–
Sundasalangidae	4	2.6	0.9	0.1	0.2	-4.5
Symphysanodontidae	4	16.5	2.8	0.2	-0.4	1.0
Synanceiidae	28	18.5	2.8	0.5	0.8	0.7
Synphobranchidae	25	66.1	4.0	0.6	-0.1	-0.8
Synbranchidae	13	56.6	3.8	0.8	-0.3	-0.5
Syngnathidae	258	16.3	2.6	0.7	-0.1	0.1
Synodontidae	57	31.7	3.3	0.6	-0.1	-0.5
Telmatherinidae	17	8.4	2.0	0.4	1.0	0.3
Terapontidae	43	22.9	3.0	0.4	0.1	-1.0
Tetrabrachiidae	1	7.0	1.9	–	–	–
Tetragonuridae	2	60.0	4.1	0.2	–	–
Tetraodontidae	155	24.2	2.9	0.7	0.1	-0.3
Tetrarogidae	33	13.9	2.4	0.6	0.8	1.9
Thaumatichthyidae	5	9.9	2.3	0.2	0.4	-1.4
Torpedinidae	14	86.6	4.3	0.6	-1.2	2.3
Toxotidae	5	23.0	3.0	0.5	0.8	-2.1
Trachichthyidae	31	23.4	2.9	0.7	0.6	-0.4
Trachinidae	8	31.0	3.3	0.6	0.1	-2.4
Trachipteridae	10	187.8	5.2	0.4	0.3	-1.9
Triacanthidae	7	24.0	3.2	0.3	-0.7	-0.5
Triacanthodidae	16	12.1	2.4	0.4	-0.5	0.7
Triakidae	36	130.1	4.8	0.4	-0.5	-0.8
Trichiuridae	39	97.7	4.4	0.6	-0.1	0.1
Trichodontidae	9	18.6	2.9	0.3	0.4	0.2
Trichomycteridae	165	7.8	1.9	0.6	0.2	0.1
Triglidae	94	23.9	3.0	0.5	0.5	-0.1
Triodontidae	1	54.0	4.0	–	–	–
Tripterygiidae	138	4.9	1.5	0.5	0.8	0.4
Umbridae	5	17.4	2.8	0.5	0.4	1.7
Uranoscopidae	39	45.5	3.3	1.1	-0.5	1.4
Urolophidae	38	42.3	3.7	0.4	-0.7	1.1
Valenciidae	2	8.0	2.1	0.0	–	–
Velliferidae	1	28.0	3.3	–	–	–
Xenisthmidae	5	3.0	1.1	0.2	1.1	1.1
Xiphiidae	1	455.0	6.1	–	–	–
Zanclidae	1	23.0	3.1	–	–	–
Zaprionidae	1	88.0	4.5	–	–	–
Zeidae	4	82.5	4.4	0.1	-1.0	-0.7
Zenionidae	6	11.0	2.4	0.3	-0.1	1.5
Zoarcidae	210	25.6	3.1	0.5	0.2	-0.3
Count	24259					
Min	1	1.6	0.4	0.0	-2.2	-5.6
Max	1895	2000	7.6	3.6	2.2	6.4
Average	47.2	57.8	3.3	0.5	0.0	0.1